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REVIEW

DRUG LEVEL MONITORING: CHROMATOGRAPHY OF SOME MINOR GROUPS OF DRUGS

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INTRODUCTION

It is obvious at first sight that the previous chapters of this volume are far from being complete with regard to different types of drugs that are accessible to monitoring by chromatographic methods or to pharmacokinetic studies. In the present chapter we have tried to accumulate additional information about some minor groups of drugs that were not worth a separate chapter although their analysis appears quite important from the pharmacological point of view. Even with this chapter included we are aware of the fact that readers may not find the drug of their interest within this volume because of the vastness of the field which is difficult to fit into a single volume. Another reason is the speed with which this area is expanding and also that we have had to cut the available literature information as we had to keep a reasonable limit on the contributions. The present survey covers the period from January 1978 to June 1984. As a basis for this survey we have used the bibliography section of the *Journal of Chromatography* and occasionally we have completed the references found herein by additional ones from our own experience. These are the references which from the viewpoint of the *Journal of Chromatography* bibliography section seemed less important and were purposely deleted at this stage.

We have attempted to summarize the facts that in our opinion are the important ones about a particular drug and which can help the reader in a rapid search for information and possibly for a choice for future experiments of his own. The table form presentation of the data appeared to us the only way how to handle the large amount of available information. In the tables it is possible to find the chromatographic techniques used together with the specification of the stationary and mobile phases, the detection procedures used and the more important parameters of the instrumental techniques. Particular attention is paid to injector, column and detector temperatures, carrier gas and mobile phase flow-rates and gradient set-up. In all cases the source of material for analysis is also specified. Where data were available, the sensitivity limit is also given. It seems to us that the information available from the tables is sufficient for planning and managing new experiments. There are, however, some entries that are incomplete. These are mainly those in which the original papers were not available to us and which we were made aware of through *Chemical Abstracts*.

The drugs are categorized according to *The Merck Index* (10th ed., Merck, Rahway, NJ, 1983) and M. Negwer (*Organisch-chemische Arzneimitteln und ihre Synonyma*, 5th ed., Akademie Verlag, Berlin, 1978) and listed alphabetically. Individual drugs in their vast majority are listed according to their generic names (if available). In those situations where there are commonly used synonyms, these are introduced as a cross-reference or listed in parentheses after the generic name. It should be pointed out, however, that not

all synonyms were considered in this respect. In any case we have avoided chemical names wherever possible.

In those instances where a particular drug can be categorized in two or more different ways, we have attempted to list it under the most common group. In some instances drugs are mentioned that have already occurred in individual chapters of this volume, clearly because of the possibility of multiple therapeutic applications. For complete information the reader is directed to the Index of Compounds Chromatographed.

Also, even at an early stage of compiling the data it became evident that not all metabolites could be specified within the survey as in many instances their chemical structure remains obscure or is only partly solved. In those cases where mixtures of drugs are subjected to chromatography, it was technically not feasible to list all of them in the form of cross-references. Readers are therefore advised to check the whole table relevant to a particular category of drugs for completeness.

TABLE 1

ANTHELMINTICS								
Drugs separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Albendazole + metabolites	TLC	Silica gel	Chloroform-methanol (9:1)	UV or autoradiography	Urine	Urine		1
	TLC	Silica gel	Chloroform-diethyl ether-acetic acid (6:1:1)	UV or autoradiography	Urine	Urine		1
	TLC	Silica gel	Chloroform-methanol-conc. ammonia (90:10:1)	UV or autoradiography	Urine	Urine		1
Dichlorvos, trichlorfon	GC-MS GC-MS			Blood, tissues	Rearrangement products of metronidazole also separated			2
Diethylcarbamazine (Banocide, Hetrazan)	GC	2% Carbowax 20 M, 5% KOH on Chromosorb G AW DMCS, 100-120 mesh	Nitrogen, 40 ml/min	Nitrogen-sensitive detection	Below 1 ng/ml	Plasma, urine	Column 160°C; injector 180°C, detector 240°C	3
Levamisole	GC	3% OV-17 on Supelcoport, 80-100 mesh, silylated column	Nitrogen, 55 ml/min	FID (rubidium sulphate)	4 ng	Plasma	Injector 255°C, oven 250°C, detector 260°C	5
Mebendazole	HPLC	μ Bondapak C ₁₈	KH ₂ PO ₄ -NaOH buffer pH 6.0-acetonitrile (73:27), 2.5 ml/min	UV 313 nm	10 ng/ml	Plasma		6
	HPLC	LiChrosorb Si 60 (5 μ m)	Chloroform (500 ml) in 10 l water, mixed, aqueous phase discarded	UV 307 nm	Below 20 ng/ml	Plasma		7
	HPLC	LiChrosorb Si 60	Acetonitrile-chloroform saturated with water-25% ammonia (75:92.5:0.1) pH 6-7, 0.8 ml/min	UV 307 nm	Below 20 ng/ml	Plasma		7
Mebendazole + metabolites	HPLC	LiChrosorb RP-8 (10 μ m); pre-column Corasil C ₁₈	Methanol-water (55:45) (pump A); methanol-0.05 M eq. ammonium phosphate pH 5.5 (55:45) (pump B), 1.7 ml/min	UV 254 nm	20 ng/ml	Plasma	Isocratic elution: pump A 67% of the mobile phase, pump B the balance; gradient elution: pump A supplies 100% of the mobile phase; at 7 min a current gradient begins which brings pump B up to 45% at 13.2 min	8

Oltipraz	GC	3% OV-17 on Chromosorb Q, 100–120 mesh	Nitrogen, 100 kN/m ²	FID	1.0–2.0 µg/ml	Serum, plasma	Oven 235°C, injector 250°C	9
Oxfendazole	HPLC	µBondapak C ₁₈	24.5% Acetonitrile in water, 2 ml/min	UV 254 nm	0.005 µg/g	Cow's milk	Phenothiazine sulphoxide: UV difference measured at 271/400 nm	10
Phenothiazine + metabolite (phenothiazine sulphoxide)	TLC	Silica gel	Chloroform—acetone (9:2)	Radioactivity (after elution), or UV at 254/400 nm, relative absorption difference	Urine, blood	Phenothiazine sulphoxide: UV difference measured at 271/400 nm	11	
	TLC	Silica gel	Tetrachloromethane—acetone (4:1)	Radioactivity (after elution), or UV at 254/400 nm, relative absorption difference	Urine, blood	Phenothiazine sulphoxide: UV difference measured at 271/400 nm	11	
	TLC	Silica gel	Hexane—acetone (3:2)	Radioactivity (after elution), or UV at 254/400 nm, relative absorption difference	Urine, blood	Phenothiazine sulphoxide: UV difference measured at 271/400 nm	11	
Piperazine, mono-nitroso and N,N'-di-nitroso derivatives	TLC	Silica gel	Ethyl acetate—methanol—aq. ammonia (0.88 sp. gr.) (17:2:1)	Radioactivity (after elution), or UV at 254/400 nm, relative absorption difference	Urine, blood	Phenothiazine sulphoxide: UV difference measured at 271/400 nm	11	
Praziquantel	GC	15% SE-52 on Chromosorb W HP	Helium, 38 ml/min	Thermal energy analyzer	0.2–2.0 ng/ml	Urine, gastric juice, blood	Column 190°C, injector 240°C, furnace 475°C	12
	GC	1.5% OV-3 on Valaspacer, 100–120 mesh	Helium, 30 ml/min	NPD	0.01 µg/ml	Serum, urine, faeces	Column 210°C, injector 290°C, detector 300°C	13
	HPLC	RP-18 (5 µm) Spheri-3 (identical pre-column)	38% Aq. acetonitrile, 1.5 ml/min	UV 210 nm	2.5 ng/ml	Serum		14
Thiabendazole, 6-hydroxythiabendazole	HPLC	µBondapak C ₁₈ , (10 µm); pre-column Bondapak C ₁₈ /Corasil (37–50 µm)	0.01 M Phosphate buffer pH 7.0—methanol (50:50)	Fluorescence at 305/370 nm or 470 cut-off filter	0.1 µg/ml thiabendazole; 0.4 µg/ml 5-hydroxythiabendazole	Serum		15
	HPLC	µBondapak C ₁₈ , (10 µm); pre-column Bondapak C ₁₈ /Corasil (37–50 µm)	0.01 M Phosphate buffer pH 7.0—methanol (55:45)	Fluorescence at 305/370 nm or 470 cut-off filter	0.1 µg/ml thiabendazole; 0.4 µg/ml 5-hydroxythiabendazole	Serum	Specifically for 5-hydroxy-thiabendazole	15

Trichlorfon (metrifonate), see Dichlorvos

TABLE 2
ANTIARTERIOSCLEROTICS (HYPOLIPIDEMICS)

Drugs separated	Method	Stationary phase	Mobile phases or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Bezafibrate	GC	8% OV-101 on Gas-Chrom Q, 80–100 mesh	Nitrogen, 60 ml/min	FID	0.2 µg/ml	Serum, urine	Methyl derivatives; injector, detector 200°C, oven 280°C	16
Ciprofibrate	HPLC	C ₁₈ -Phenyl (10 µm), fatty acid analysis column	Tetrahydrofuran–0.1 M K ₂ HPO ₄ buffer pH 4–aceto-nitrile (10:104:96), 2 ml/min	UV 232 nm	0.016 µg/ml	Plasma	Guard column phenyl Corasil 37–50 µm	17
Clofibrate	GC	5% SE-30 on Supelcoport, 80–100 mesh	Nitrogen, 20 ml/min	FID		Plasma	Clean-up procedure; penta-fluorobenzoyl derivatives; column programmed 150–245°C, 8°C/min	18
Clofibrate, etofibrate, clofibrate acid, clofibrate acid monoglycolate, nicotinic acid, nicotinic acid monoglycolate	HPLC	RCM 100 C ₁₈ , µBondapak alkylphenyl	Nitrogen, 18 ml/min	FID		Plasma	Pentafluorobenzoyl derivatives; column isothermal 180°C	18
	HPLC	Acetonitrile–0.1 M acetate buffer pH 3.75 (50:50), 1.5 ml/min		UV 225 nm		Plasma, urine		19
	HPLC	Acetonitrile–0.05 M acetate buffer pH 3.5 (50:50), 6 ml/min		UV 225 nm		Plasma, urine		19
	HPLC	Acetonitrile–0.1 M acetate buffer pH 3.75 (45:55), 1.5 ml/min		UV 225 nm		Plasma, urine		19
	HPLC	Methanol–0.1 M acetate buffer pH 3.75 (35:65), 1.5 ml/min		UV 225 nm		Plasma, urine		19
	HPLC	Methanol–0.05 M acetate buffer pH 3.5 (50:50), 3 ml/min		UV 225 nm		Plasma, urine		19
	HPLC	Acetonitrile–0.05 M acetate buffer pH 3.5 (50:50), 3 ml/min		UV 225 nm		Plasma, urine		19
	HPLC	Acetonitrile–0.05 M acetate buffer pH 3.5 (30:70), 3 ml/min		UV 225 nm		Plasma, urine		19

Clofibrate acid (see also Clofibrate)	GC	3% OV-17 on Chromosorb W HP, 80–100 mesh	Nitrogen, 30 ml/min	FID and UV (compounds in the effluent trapped in 2-propanol)	Serum	20	
	GC	Coiled Ni column packed with 10% EGA on Chromosorb W HP, 80–100 mesh μ Bondapak C ₁₈	Nitrogen, 30 ml/min	FID and UV (compounds in the effluent trapped in 2-propanol)	Serum	20	
	HPLC	0.01 M Citrate buffer pH 2.5—acetonitrile (65:35)	¹⁴ C Scintillation	¹⁴ C-labelled clofibric acid	Plasma	21	
	HPLC	Acetonitrile—0.5% acetic acid (42:58), 70 ml/h	UV 235 nm	0.5 μ g/ml	Plasma, urine, saliva	22	
Clofibrate acid, probenecid	HPLC	Acetonitrile—acetic acid—water (450:5:545), 2 ml/min (10 μ m)	UV 235 nm	10 μ g/ml (clofibric acid), 15 μ g/ml (probenecid)	Plasma, urine	23	
	TLC	Silica gel	UV or chemical detection	Urine	Qualitative separation only	24	
Clofibrate acid + clofibrate metabolites	TLC	Silica gel	UV or chemical detection	Urine	Qualitative separation only	24	
	TLC	Silica gel	UV or chemical detection	Urine	Qualitative separation only	24	
Gemfibrozil + metabolites	GC	3% OV-22 on 80–100 mesh Supelco GCQ (plasma), 10% Poly I:110 on 80–100 mesh Supelco GCQ (urine)	Nitrogen, 50 ml/min	FID	0.5 μ g/ml (plasma); 5 μ g/ml (urine)	Column 180°C, injector 230°C, detector 250°C	
[1-O-(<i>p</i> -Myristyloxy)- α -methylcinnamoyl]- glycerol (IK-903)	TLC	Silica gel	Light petroleum—ethanol (87.5:12.5:1)	Fluorescence at 290/350 nm	Plasma	After first fluorimetry redevel- opment in light petroleum— acetone-acetic acid (90:10:15:1) and second fluorimetry	
Parmidine	TLC	Silica gel	Chloroform—methanol (10:1)	UV 260 nm	Serum, urine	27	
Probucool	HPLC	μ Bondapak C ₁₈ , (10 μ m)	Acetonitrile—water (86:15), 2 ml/min	UV 254 nm	0.25 μ g/ml	Plasma	28

TABLE 3
ANTIBACTERIALS (INCLUDING ANTISEPTICS, DISINFECTANTS, CHEMOTHERAPEUTICS)

Cross-references between individual mixtures of sulpha drugs are not supplied. Check the whole table for completeness.

Drugs separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
<i>p</i> -Aminosalicylic acid	HPLC	LiChrosorb C ₁₈ RP (10 µm)	Methanol—water (20:80) cont. hydroxide and 0.01 M disodium acid phosphate pH 5.5, 1.0 ml/min	Fluorescence at 270/385 nm	500 µg	Plasma		29
Chlorhexidine (Hibitane)	HPLC	ODS Waters Assoc. RP (10 µm)	1000 µg/ml Toluene-4-sulphonic UV 238 nm acid in methanol—water (65:35), 1.5 ml/min	0.2 µg/ml	Blood, urine			30
	HPLC	µBondapak C ₁₈ (10 µm)	Methanol—20 mM sodium acetate buffer pH 5 (60:40) + 100 µg/ml pentadecafluoro-octanoic acid, 1.5 ml/min	UV 260 nm	0.1 µg/ml	Urine	Extraction from urine using Sep-Pak	31
Chinoxacin + metabolites	HPLC			UV 254 nm	0.5 ng/ml	Urine		32
Clofazamine (Lamprene)	HPLC	RP-UltraspHERE-octyl (5 µm), 40°C	0.0425 M Phosphoric acid in 81% methanol pH 2.4, 1.5 ml/min	UV 285 nm	10 ng/ml	Plasma	Combination with MS	33
	HPTLC	Silica gel	Predevelopment with chloroform—methanol (1:1), then sampling and development with toluene—acetic acid—water (50:5:3:4)	Densitometry at 545 nm	5 ng/g	Plasma		34
Clofazamine + metabolites	TLC	Silica gel	Butanol—benzene—water—methanol (2:1:1:1:25), then elution from the plate and purification with HPLC	UV		Urine	Combination with HPLC	35
	HPLC	Altex Ultrasphere ODS RP	60% Methanol—water (first metabolite); 55% methanol—water (second metabolite)			Urine	Combination with HPLC	35

Cotrimoxazole, see Trimethoprim

Cyclindole + metabolites	GC	10% OV-1 on Gas-Chrom Q, 100–120 mesh	Nitrogen, 45 ml/min	FID	Below 10 ng/ml	Blood, urine	^{14}C ; column 230°C, flash heater and detector 250°C; trimethylsilylimidazole derivatives	36
Dapsone (DDS, 4,4'-diaminodiphenylsulphone) + mono-acetyl derivative	HPLC	μ -Bondapak C ₁₈ (10 μm)	Acetonitrile–1.5% acetic acid (26:74), 2 ml/min	UV 280 nm	Below 0.2 ng/ml	Serum	Switching of the mobile phase is performed	37
Dapsone, clofazamine, rifampicin	HPLC	μ -Bondapak C ₁₈	(A) Acetonitrile–water (20:80), 2 ml/min; (B) Tetrahydrofuran–0.5% acetic acid (40:60), 1.5 ml/min; (C) Tetrahydrofuran–water (50:50), cont. 0.025 M 1-pentanesulphonic acid in glacial acetic acid, 1.5 ml/min	UV 261, 296 nm for dapsone; 287 nm for clofazamine, 242, 266, 337 nm for rifampicin	Below 0.2 ng/ml	Serum	Switching of the mobile phase is performed	38
2,4-Diamino-5-[3,5-dimethoxy-4-(methylthio)benzyl]pyrimidine (Ro-12-6395)	HPLC	LiChrosorb Si 60 (5 μm)	5 ml Methanol and 0.3 ml 33% aq. ammonia diluted to 100 ml with ethyl acetate	Fluorescence at 290/340 nm	0.02 $\mu\text{g}/\text{ml}$	Plasma	Column 150°C, injector 180°C, separator 225°C	39
Ethambutol	GC–MS	2% OV-17 on Gas-Chrom Q, 80–100 mesh	Helium, 40 ml/min	SIM	Below 0.1 $\mu\text{g}/\text{ml}$	Plasma	Column 150°C, injector 180°C, separator 225°C	40
	GC–MS	3% OV-17 on Gas-Chrom Q, 100–120 mesh	Helium, 20 ml/min	MS		Plasma	Column 160°C, injector 190°C, jet separator 200°C, ion-source 250°C	41
	GC	3% SE-30 on Gas-Chrom Q, 100–120 mesh	Nitrogen, 30 ml/min	^{63}Ni ECD	36 ng/ml	Plasma	Column 170°C, injector 190°C, detector 250°C	41
	GC	OV-101 on Gas-Chrom Q, 100–120 mesh	Nitrogen, 20 ml/min	^{63}Ni ECD		Plasma, urine	Column 155°C, injector 210°C, detector 240°C; derivatization with trifluoroacetic anhydride	42
Ethionamide, prothionamide	HPLC	μ -Porasil (10 μm)	Diethyl ether–methanol (96:4), UV 295 nm 1.3 ml/min	0.01 $\mu\text{g}/\text{ml}$		Plasma, serum urine	(Continued on p. 410)	43

TABLE 3 (continued)

Drugs separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
1-Ethyl-5-fluoro-1,4-dihydro-4-oxo-7-(1-piperazinyl)-1,8-naphthyridine-3-carboxylic acid (AT-2266)	HPLC	μ Bondapak C ₁₈ (10 μ m)	Methanol—0.1 M citric acid—acetonitrile (9:5:1 or 6:5:1); methanol—5% acetic acid—acetonitrile (6:10:1)	UV 340 nm	0.01 μ g/ml (plasma); 0.1 μ g/ml urine	Plasma, urine		44
4-Ethylsulfonylnaphthalene-1-sulphonamide + metabolites	HPLC	Vydac RP (10 μ m); μ Bondapak C ₁₈ (10 μ m); Zorbax ODS (5–7 μ m)	43% Methanol in 0.02 M ammonium acetate, 0.5 ml/min; isopropanol in 0.02 M ammonium acetate, gradient from 1% to 10% over 30 min, 0.5 ml/min	(1) UV 254 or (2) fluorescence 310/390 nm		Urine		45
Fenclofene	GC	3% SE-30 on Chromosorb	Argon—methane (95:5), 50 ml/min	⁶³ Ni ECD	0.3 μ l/ml	Plasma	Derivatization with diazo-methane	46
Furazolidone	HPLC	Pre-column Perisorb RP-8 (30–40 μ m); analytical column Hyperal SAS Partial PXS 10/25 ODS	Water—acetonitrile (75:25), 1.75 ml/min	UV 360 nm	0.05 mg/kg	Liver, kidney		47
Iodochlorhydroxyquin (cloquinol, Chinoform, Vioform)	HPLC		Methanol—water (30:70), buffered to pH 4.0 with dibasic sodium phosphate (16.5 mM) and sodium citrate (13.1 mM), 1.1 ml/min	UV, electrochemical detection	1 ng/ml	Plasma, serum		48
	GC	3% OV-101 on Chromosorb W HP, 80–100 mesh	Argon—methane (9:1), 60 ml/min	ECD	50 ng/ml	Plasma	Column 195°C, injector 250°C, detector 300°C	49
	GC-MS	1% OV-17 on Chromosorb W AW DMCS, 80–100 mesh	Helium, 37 ml/min	Multiple-ion detection	1 ng	Plasma	Separation after acetylation; column, injector 215°C, separator 210°C, ion-source 200°C	51
	RP-HPLC	ODS-HC-SIL-X-1 (10 μ m); guard column Rheodyne RP-18-MPLC	Methanol—0.05 M phosphoric acid (70:30), 1 ml/min	UV 256 nm	0.2 μ g	Urine, faeces, liver		52
	HPLC	Guard column ODS-HC-SIL-X-1; RP-18-MPLC	Methanol—0.05 M phosphoric acid (80:20)	UV 256 nm	Below 1 μ g/ml	Plasma		53

Ioniozaid + its conjugate	TLC	Silica gel	<i>n</i> -Butanol—acetone—diethylamine—water (30:20:4:30)	UV 267 nm	0.04 µg/ml	Plasma	54
Ioniozaid + metabolites (acetyl-hydrazine, diacetyl-hydrazine and acetylisoniazid)	GC—MS	1% OV-17	Helium, 30 ml/min	MS	0.01—2 µg/ml for metabolites	Trimethylsilyl derivatives of acetylisoniazid and diacetyl-hydrazine; benzaldehyde hydrazones of ioniazid and acetylhydrazine; column 90—270°C, programmed heating (10°C/min)	55
	GC—MS	1.6% OV-17 on Shimalite W, 80—100 mesh	Helium, 30 ml/min	MS	0.01—2 µg/ml for metabolites	metabolites acetylhydrazine and diacetylhydrazine; column 90—270°C, programmed heating (10°C/min)	55
	GC—MS	RP Ultrasphere-octyl (5 µm), 40°C	0.0425 M Phosphoric acid in 81% methanol pH 2.4, 1.6 ml/min	UV 265 nm	10 ng/ml	Rat hepatocytes	56
Ioniozaid + metabolite (acetylisoniazid), rifampicin + metabolite (deacetyl-rifampicin)	HPLC	µBondapak C ₁₈ (1.0 µm)	Methanol—water (5:95) cont. 5 mM <i>n</i> -heptanesulphonic acid, 2 ml/min	UV 260 nm	95 ng/ml	Serum, polymorphonucleocytes, alveolar macrophages	57
	HPLC	Zorbax CN RP (6 µm)	Methanol—water—acetic acid (40:5:90:1.0) cont. 3 mM CH ₃ COONa · 3H ₂ O	UV 330 nm	1.7 ng/ml	Plasma, serum	58
	HPLC	Silica (5—7 µm)	Chloroform—methanol—water—ammonium hydroxide (84:22:1.6:1.68±0.2), 0.5 ml/min 0.01 M Phosphoric acid in acetonitrile—water (20:80), 2 ml/min	UV 254 nm	50 ng/ml	Plasma	59
	HPLC	Spherisorb nitrile (5 µm)		UV 266 nm	0.02 µg/ml	Plasma	60
Ioniozaid (A), rifampicin (B), acetylisoniazid (C)	HPLC	RP-18	Methanol—1% triethanolamine (36:15:1.6:1.68±0.2), 0.5 ml/min 0.01 M Phosphoric acid in acetonitrile—water (20:80), 2 ml/min	UV 260 nm	0.5 µg A; 1.0 µg B and C	Serum	61
Mandelic acid	GC	(A) 3% OV-1; (B) 3% OV-17; or (C) 3% SP-1000 on Chromosorb W HP, 100—120 mesh	Nitrogen, 20 ml/min	FID	1 µg/ml	Plasma	62
					Determination of butyl esters; column: (A) 1120°C, (B) 140°C, (C) 170°C, injector and detector temperatures 30°C higher		63

(continued on p. 412)

TABLE 3 (continued)

Drugs separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Methenamine	GC	OV-17 capillary column	Nitrogen, 1.0–1.2 ml/min	Nitrogen-sensitive detection		Plasma, urine	Injector 240°C, oven 150°C, detector 250°C	65
Nalidixic acid	GC	10% OV-17 on Chromosorb W HP, 80–100 mesh	Nitrogen, 20 ml/min	FID	0.1–0.3 µg/ml	Plasma	Column 270°C, injector 280°C, detector 290°C; ifufenamic acid was added to deactivate the column	66
	GC	(A) 3% OV-1; (B) 3% OV-17; or (C) 3% SP-1000 on Chromosorb W HP, 100–120 mesh Partisil PXS 10/25 PAC	Nitrogen, 20 ml/min	FID	1 µg/ml	Plasma	Determination of butyl ester; column: (A) 230°C, (B) 250°C, (C) 270°C, injector and detector temperatures 30°C higher	64
	HPLC	(A) µBondapak C ₁₈ ; (B) LiChrosorb 10 RP-18 C ₁₈ (10 µm)	Methanol—0.1 M citrate buffer pH 3 (95:15), 1.6 ml/min	UV 254 nm	0.05 µg/ml (plasma); 0.42 µg/ml (urine)	Plasma, urine	Determination of main metabolite, 1-ethyl-1,4-dihydro-4-oxo-1,3-naphthyridine-3,7-dicarboxylic acid	67
	HPLC	(A) µBondapak C ₁₈ ; (B) LiChrosorb 10 RP-18 C ₁₈ (10 µm)	65% Methanol in water for A; 70% Methanol in water for B, 1.5 ml/min	UV 313 nm	1.0 ng per injection	Plasma		68
Nalidixic acid + metabolites (7-hydroxynalidixic acid and 7-carboxynalidixic acid)	HPLC	RP-8 Hewlett-Packard (10 µm)	Methanol—phosphate buffer pH 8.2 (55:45) cont. 2 g/l N,N,N-trimethylacetyl-ammonium bromide	UV 254 nm	0.5 µg/ml	Plasma, urine		69
Nalidixic acid + metabolite (hydroxy-nalidixic acid)	HPLC	µBondapak C ₁₈ (10 µm)	Water—methanol (1:1) cont. phosphate buffer pH 7.4; final concentration is 0.016 mol/kg phosphate and 0.12% (w/w) ceftriaxone, 1.5 ml/min	UV 313 nm	1 µg/ml			70
	TLC	Silica gel	Dioxane—5 × diluted conc. ammonia (3:1)	Fluorescence at 375/430 nm	0.16 µg/ml		Hydroxynalidixic acid may interfere	71
Nitrofurantoin, Urafadyn (hydroxymethyl furantoin)	HPLC	LiChrosorb 5 RP-8 (5 µm)	Water with 5% ethanol, 1.6 ml/min	UV 370 nm	0.02 µg/ml	Plasma, urine		72
Nitrofurantoin	HPLC	LiChrosorb C ₁₈ RP	Methanol—0.01 M acetic acid (2.5:97.5), 2 ml/min	UV 280, 365 nm		Plasma, urine		73
	HPLC	µBondapak C ₁₈	Water—methanol (100:10), gradient to 70:30 in 20 min, 4 ml/min	UV 280 nm		Liver, kidney, lung, small intestine walls, ileum, colon contents	Metabolite M-4 UV 365 nm	74

HPLC	μ Bondapak C ₁₈ , RP	Methanol—0.01 M sodium acetate buffer pH 5.0 (20:80) Methanol—water (50:50), 1 ml/min	UV 365 nm UV 368–371 nm	Below 0.02 μ g/ml 0.2 μ g/ml (HPLC); 0.4 μ g/ml (polarography)	Plasma, urine Serum	75	
HPLC	LiChrosorb				Combination of HPLC and differential pulse polarography	76	
HPLC	Knauf Fertigsaule RP-2 35°C	Acetonitrile—water—85% phosphoric acid (20:80:0.5), 2.2 ml/min	UV 380 nm	Below 10 μ g/ml	Serum, urine	77	
Nitrofurantoin, Urfadyn HPLC (hydroxymethyl-furanone)	μ Bondapak C ₁₈	30% or 40% methanol cont. 0.5% glacial acetic acid	UV 365 nm		Urine	78	
Pefloxacin + metabolites (pefloxacin N-oxide, desmethyldifloxacin, norfloxacin)	HPLC	LiChrosorb RP-18 (10 μ m)	(A) Dist. water; (B) acetonitrile—water (2:3). For pefloxacin 5.2% and 48% B; for metabolites gradient starting from 0–20% B and rising at a rate 2.5% per 10 min, 2 ml/min	UV 270 nm	0.05 μ g/ml (plasma); 0.5 μ g/ml (urine)	Mixture of acetonitrile and water contains 0.4%; Na ₂ HPO ₄ • 12H ₂ O and 0.2% tetraethylammonium iodide pH 9.4	79
Piperimic acid	HPLC	μ Bondapak C ₁₈ , RP (10 μ m); guard column Co-Pell ODS C ₁₈ , (30 μ m)	Serum: 46.8 g NaH ₂ PO ₄ • 2H ₂ O UV 280 nm in 75 ml methanol, 75 ml acetonitrile and 850 ml water; urine: 46.8 g NaH ₂ PO ₄ • 2H ₂ O in 275 ml methanol and 725 ml water	0.1 μ g/ml (serum); 5 μ g/ml (urine)	Serum, urine	80	
Protonamide, see Ethionamide							
Pyrazinamide + metabolites (5-hydroxypyrazine-2-carboxylic acid, pyrazinoic acid)	GC-MS	3% OV-17 on Chromosorb W HP, 80–100 mesh	Isobutane	MS	20 ng/ml	Silyl derivatives; column programmed 120–180°C (15°C/min), then 250°C; no separator between GC and MS; connecting tube 240°C, inlet 250°C	81

(Continued on p. 414)

TABLE 3 (continued)

Drugs separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Resoracin + N-oxide metabolite	HPLC	Partisil-PXS 10/25 PAC; pre-column Corasil	Acetonitrile—0.2 M phosphoric acid (98:2), 2 ml/min	UV 280 nm	0.13 µg/ml resoracin in plasma, 0.64 µg/ml resoracin in urine, 0.21 µg/ml metabolite in plasma; 0.60 µg/ml metabolite in urine	Plasma, urine		82
Rifampicin, see Dapsone, Isoniazid and chapter on Antibiotics								
Seddamine	TLC	Silica gel	Benzene—acetic acid (90:30)	UV 254 nm or ¹⁴ C liquid scintillation counting		Urine		83
Salicylic acid	GC	10 µm Methyl-silicone-coated fused-silica capillary column	Helium, 1.2 ml/min	FID		Urine	Silyl derivatives; injector and splitter 250°C, oven programmed 60–200°C (8°C/min), detector 250°C	84
Salicylic acid, salicyluric acid	HPLC	µBondapak C ₁₈ (10 µm)	Methanol—water—glacial acetic acid (50:60:1), 2 ml/min	Fluorescence at 240/350 nm	150 ng/ml salicylic acid; 300 ng/ml salicyluric acid	Plasma, urine		85
SC-38538, i.e. sodium 4-(2-(1-methyl-5-nitro-imidazolylthio)-ethoxy)benzoate	LC	ODS Hypersil (5 µm)	50% Methanol—0.01 M K ₂ HPO ₄ pH 3.5, 2 ml/min	UV 254 nm	0.05 µg/ml	Plasma, urine		86
Sulphadiazine + sulphathiazole, sulphamerazine, sulphapyridine	HPLC	LiChrosorb RP-8	Plasma: pH 5 buffer—acetone nitrile (80:1); 40°C, 2 ml/min urine: pH 4 buffer—acetone nitrile (92:8), 40°C, 2.5 ml/min Co: Pell ODS	UV 254 nm	250 ng/ml (plasma); 2.5 µg/ml (urine)	Plasma, urine		87

Sulphadiazine, sulphamerazine, sulphamerazine	HPLC	LiChrosorb RP-C ₁₈ (10 μm)	400 ml Methanol + 600 ml water + 1.6 g lithium perchlorate	Electrochemical detection, ampero- metric determination	10 ng/g	Liver, kidney, muscle tissues	88
Sulphadiazine, sulphadimethoxine, sulphamerazine, sulphisoxazole	HPLC	LiChrosorb RP-C ₁₈ (10 μm)	Methanol—water (25:75 or 40:60) with 0.01 M lithium perchlorate	Electrochemical detection	10 ng/ml	Milk	89
Sulphadiazine, sulphadimethoxine, sulphamerazine, sulphisoxazole	LC—MS	Whatman PXS 10/25 ODS	Linear gradient acetonitrile— water (10:90 to 90:10) over 10 min	UV 254 nm, detector connected with Hewlett-Packard DIL LC-MS system	Low nano- gram range	Plasma, urine	90
Sulphadiazine, sulphamerazine, sulphamerazine, N-acetylsulphameth- oxazole	HPLC	μBondapak/C ₁₈ RP ODC		UV 254 nm		Plasma	91
Sulphadiazine, sulphamerazine, sulphamerazine, N-acetylsulphameth- oxazole	TLC	Silica gel	Toluene—isopropanol (8:2); chloroform—ethanol (8:1 or 8:2)	Densitometry 575 nm	2.5 μg/ml (plasma); 10 μg/ml (urine)	Urine, plasma	92
Sulphadiazine, sulphadimethoxine, sulphisomidine	TLC				0.5 μg per spot	Plasma	93
Sulphadiazine, sulphamerazine	GC, TLC		5% OV-61 on Gas-Chrom Q _{80–100} mesh	⁶³ Ni ECD	50 pg/ml	Urine	Column 260°C, injector 270°C, detector 350°C
	HPLC	LiChrosorb 10 RP 18	Acetonitrile—10 mM acetate buffer pH 4.0 (1:9), 2.0 ml/min	UV 254 nm		Fat, kidney, liver, porcine muscle Plasma	94
							95
							96

(Continued on p. 416)

TABLE 3 (continued)

Drugs separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Sulphamethazine + polar metabolite	HPLC	μ -Bondapak C ₁₈	(A) Methanol-water (5:95); (B) acetonitrile-isopropanol—water (5:0.1:94.5); (C) methanol—water (30:70);— 2 ml/min	UV 250 nm	Porcine liver	Connected with MS		97
Sulphamethazine + metabolite	TLC	Silica gel	Benzene-methyl ethyl ketone —ethanol—water (30:30:30) Benzene-methyl ethyl ketone —ethanol—water (30:30:30) scintillation	¹⁴ C Liquid scintillation	Porcine liver	Metabolite eluted and purified with HPLC (see above)		97
Sulphamethoxazole + N-acetyl metabolite	TLC	Silica gel	(A) LiChrosorb Si 100 C ₁₈ , RP (10 μ m); (B) LiChrosorb C ₄ , RP (7 μ m)	(A) Methanol—water (3:7); UV 254 nm (B) acetonitrile—water (1:3) Acetonitrile—5 mM aq. acetic acid (1:3); 1.0 ml/min.	2 μ g/ml	Urine	Live, kidney, muscle, fat	98
Sulphamethoxazole + metabolites	HPLC	μ -Bondapak CN	RP (10 μ m)	UV 254 nm	2 μ g/ml	Urine		99
Sulphamethoxazole + metabolites	LC	LiChrosorb RP-8 (10 μ m), 65°C	FID	50 ng	Urine	Pyrolyser connected to GC dual columns; pyrolysis 770°C 5 sec., injector 275°C, detector 350°C, column programmed from 100°C to 245°C (5°C/min)		100
Sulphamethoxydiazine + metabolites	GC-MS, pyrolysis	8% Carbowax 20 M and 2% KOH on Chromosorb W AW DMCS, 100-120 mesh	Helium or nitrogen, 50 ml/min	50 ng	Urine	Pyrolyser connected to GC dual columns; pyrolysis 770°C 5 sec., injector 275°C, detector 350°C, column programmed from 100°C to 245°C (5°C/min)		101
Sulphamethoxy-pyridazine	HPTLC	Silica gel	Chloroform-acetic acid—methanol (18:2:1:0:0.8)	Fluorescence at 366 nm	Plasma			102
Sulphapyridine	HPLC	RP-18 (5 μ m)	0.05 M Na ₂ HPO ₄ , 0.01 M 1-hexanesulphonate, sodium salt, 0.0072 M triethylamine and 15% methanol, 1.0 ml/min	Fluorescence at 395/470 nm	5 ng/ml	Saliva	On-line derivatization with fluorescamine	103
Sulphapyridine + N-acetyl derivative	HPLC	μ -Bondapak CN	Ag 0.4% sodium acetate and 4% acetic acid, 2.0-2.3 ml/min	UV 254 nm	0.25 μ g/ml	Plasma, saliva	Metabolites of propxyphenine may interfere	104
GC, HPLC	LC	Silica	Chloroform—acetonitrile—methanol—35% ammonia (65:5:30:4:0.5), 1 ml/min		5 ng	Plasma, saliva, urine		105

Sulphapyridine, sulphaquazidine, sulphamerazine, sulphamethazine, sulphanilamide, sulphathiazole, sulphisoxazole, acetyl- sulphisorezole	HPLC	Water—methanol (60:40) adjusted to pH 4.0 with acetate buffer	UV 254 nm	10 ng/ml	Plasma	107	
Sulphapyridine, sulphamerazine, sulphamethazine, sulphathiazole, sulphisoxazole	HPLC	μ Bondapak RP C ₁₈ , 0.07 M KH ₂ PO ₄ —0.07 M Na ₂ HPO ₄ —ethanol—methanol (780:200:100:100)	UV 254 nm	1 μ g/ml	Serum	108	
Sulphaquinoxaline	HPLC	Spherisorb S10 W (10 μ m)	n-Hexane—1-hexene saturated with water—chloroform—aceto- nitrile—methanol—25% ammonia (36:30:15:4.5:4.5:0.05 or 37:30:15:1.4:5.3:5:0.05)	UV 254 nm	Below 10 ppb	Liver, muscle	109
Sulphisoxazole + N-acetyl metabolite	HPLC	Pre-column Corasil	Water—methanol (60:40) adjusted to pH 4.0 with acetate buffer	UV 254 nm	10 ng/ml	Plasma, urine	110
	HPLC	LiChrosorb RP-18 Hibar II RP	Methanol—0.01 M sodium acetate (see Note) pH 4.7 (32:68), 1.2 ml/min	UV 254 nm	Below 0.05 μ g/ml	Plasma, urine	111,
Sulphasalazine + metabolite (sulphapyridine)	HPLC	NucleoSil RP-18 (10 μ m)	50% Methanol cont. 1 g/l N,N,N-trimethyl ethyl ammonium bromide, 1.3 ml/min	Fluorescence at 310/430 nm	0.25 μ g/ml	Plasma	113, 114
	HPLC	Hyperai-SAS (5 μ m)	22.5% Methanol and 0.05 M phosphate buffer cont. 0.1% tetrabutylammonium hydrogen sulphate, 1 ml/min	UV 290 nm, coupled with fluorescence at 320/389 nm		Plasma	115
	HPLC	LiChrosorb RP-18 (5 μ m); guard-column Chrompack	Water adjusted to pH 3.3 with 0.01 M citric acid and 20% methanol, 1.4 ml/min	Fluorescence at 305 nm, cut-off filter 395 nm	0.1 μ mol/l	Serum, urine, faeces	116
Sulphasalazine + metabolite (5-amino- salicylic acid, N-acetyl- 5-amino-salicylic acid, sulphapyridine, N- acetyl sulphapyridine	HPLC	Hyperai-SAS (5 μ m)	Methanol—0.05 M phosphate buffer pH 7.4 (15:35), 1 ml/min	Fluorescence at 310/410 nm, UV 260 nm	0.05 mg/l p- aminosalicylic acid; 0.06 mg/l N-acetyl de- rivative (fluorescence detection)	117	

(Continued on p. 418)

TABLE 3 (continued)

Drugs separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Sulphasalazine (A) + metabolite [sulphapyridine (B)]	HPLC	(A) LiChrosorb Si 60 (5 µm); (B) µBondapak / Coral	For A: chloroform—methanol—35% ammonia (96.5:4:0.5); for B: 0.01 M phosphate buffer pH 7.7 with 17% acetonitrile, 1 ml/min	UV 365 nm 700 pg/ml A; 5 ng/ml B	Plasma			118
Tauroulin + metabolite (1,1-dioxoperhydro-1,2,4-thiadiazine, DPT)	HPLC	Hyperil (5 µm)	Chloroform—heptane (40:60) for DPT; methanol—chloroform fluorescence (2:98) for DNS taurine amide, 1 ml/min	UV 254 nm or Below 5 µg/ml DPT	Serum	Dns derivatives		119
Tetroxoprim (TXP), sulphadiazine	HPLC	LiChrosorb RP 8 (10 µm) for urine	Acetonitrile—water pH 7.8 (alkalized with ammonium carbonate), concentration of acetonitrile programmed from 5% to 95% (1.2 min), 3 ml/min Methanol—water (60:50) adjusted to pH 4.6 with acetic acid for tetroxoprim; acetonitrile—1% acetic acid (16:9:4) for sulphadiazine	UV 254 nm 50 ng/ml	Urine, serum	tetroxoprim; 150 µg/ml sulphadiazine		120
Tetroxoprim (TXP), sulphadiazine (SDZ), methioprim (MTP)	HPLC	ODS Hypersil (5 µm)	TXP and SDZ: 800 ml 0.1 M KH ₂ PO ₄ , cont. 1% acetic acid, 1% ethyl acetate and 200 ml acetonitrile, 1.3 ml/min; MTP and SDZ: the same mobile phase, 1.3 ml/min for 3 min, then 2.3 ml/min	UV 254 nm 50 ng/ml	Serum, TXP and prostatic secret			121
	TLC	Silica gel	Chloroform—methanol—methyl ethyl ketone—ammonia (60:22:10:4); chloroform—n-propano—formic acid (4:4:2); n-butanol—water—acetic acid (4:3:1)	¹⁴ C Autoradiography		SDZ ng/ml SDZ	Urine, plasma	122
Tetroxoprim, trimethoprim	TLC	Silica gel silanized	0.3 M sodium chloride—acetone—10 M acetic acid (100:50:0.5)	Fluorescence densitometry	50 ng/ml		Plasma	123

Thiacetazone	HPLC	μ Bondapak RP (10 μ m); pre-column Co-Pell ODS	Acetonitrile—water (3:7), 1.5 ml/min	UV 328 nm	3 ng/ml	Plasma, urine	124
Triclocarban (3,4,4'-trichloro-carbamide)	HPLC	μ Bondapak C ₁₈ RP	Water—acetonitrile (66:34), gradient to 30:70, 2.0 ml/min	UV 268 nm		Urine, plasma, Bile	125
	HPLC	μ Bondapak C ₁₈ RP	Water—acetonitrile (66:34), gradient to 30:70, 2.0 ml/min	UV 258 nm			126
Trimethoprim	GC	10% Poly S-179 on Chromosorb W HP, 80–100 mesh Porasil (10 μ m)	Helium, 45 ml/min	NPD	0.1 μ g/ml	Plasma, urine	Column 330° C, injector and manifold 350° C
	HPLC	Chloroform and a mixture methanol—water—ammonia (150:9:1) (50:0:25), 1.5 ml/min		UV 258 nm	0.01–0.02 μ g/ml	Plasma, blood, urine	127
	HPLC	Nucleosil C ₁₈ (5 μ m)	0.07 M KH ₂ PO ₄ , pH 4.75—methanol (3:1), 1.5 ml/min	UV 280 nm or electrochemical detection	0.1 ppm (UV or 0.01 ppm (electrochemical detection)	Plasma, urine	128
Trimethoprim + metabolites	HPLC	LiChrosorb RP-18 (5 μ m)	0.1 M KH ₂ PO ₄ buffer pH 7.5—acetonitrile cont. 0.7 · 10 ⁻³ M tetrabutylammonium hydrogen sulphate (85:15), 1.0 ml/min	UV 264 nm, connected with electrochemical detector	0.1 ppm	Urine	Hydrolysis of conjugates with β -glucuronidase
	HPLC	RP-8 (1.0 μ m)	1.0 M/ml sodium borate pH 9 (35:65), 1.0 ml/min	Fluorescence at 279/370 nm	0.1 μ g/ml	Plasma, urine	130
Trimethoprim, sulphadiazine, N-acetylsulphadiazine	HPLC	LiChrosorb Si 60 (10 μ m); pre-column LiChrosorb Si 60 (25–40 μ m)	Dichloromethane—methanol—25% ammonia (80:19:1), 1.5 ml/min	UV 289 nm	0.03 μ g/ml	Serum, urine	131
							132

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TABLE 3 (continued)

Drugs separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Trimethoprim, sulphamethazine, sulphadiazine, sulphanerazine, sulphathiazole	TLC				0.1 µg/ml	Serum, plasma, blood		133
Trimethoprim, sulphamethoxazole (co-trimoxazole)	HPLC	RP Spherisorb ODS (10 µm)	Acetonitrile—aq. 0.1 M KH ₂ PO ₄ , UV 254 nm cont. 1% acetic acid and 1% ethyl acetate pH 2.5 (30:7:0), 1 ml/min; for trimethoprim: methanol—water—acetic acid (40:60:1) pH 3.2, 2 ml/min	0.1—0.2 µg/ml	Plasma			134
Trimethoprim, sulphamethoxazole, N-acetylsulpha- methoxazole	HPLC	µBondapak C ₁₈ (10 µm); 30°C for serum, room temperature for urine	Methanol—1% acetic acid (200:800)	UV 230 nm	0.1 µg/ml trimetho- prim and 1.0 µg/ml for both sulphon- amides	Serum, urine	Small peak of caffeine may interfere in serum	135
	HPLC	LiChrosorb Si-60 (5 µm); pre-column LiChroprep Si-60 (25—40 µm)	Chloroform—methanol—water— ammonia (94.5:5:0.25:0.19) (solvent A) 3 min, then solvent B, otto (79:20:1:0.15), 8 min; programming 0—3 min solvent A, 3.5 min A+B (1:1), 4—12 min solvent B, 12.5 min A+B (1:1), 13—16 min solvent A, 2 ml/min	UV 280 nm	15 ng/ml trime- thoprim; 20 ng/ml sulpha- methoxazole, 10 ng/ml acetyl sulpha- methoxazole	Plasma, urine		136
	HPLC	µBondapak C ₁₈ , RP buffer pH 3.5 (35:65), 1 ml/min	Methanol—0.067 M phosphate buffer pH 3.5 (35:65), 1 ml/min	UV 225 and 254 nm	0.05 µg/ml trimethoprim; 0.2 µg/ml sulphametho- xazole; 0.5 µg/ml acetyl derivative	Serum		137
	HPLC						Serum, urine	138

Trimethoprim, sulphamethoxazole	HPLC	Waters-RP C ₁₈ (10 μm)	11.867 g Na ₄ HPO ₄ · 2H ₂ O dissolved in 1 l water (A); 9.073 g KH ₂ PO ₄ in 1 l water; pH 5.0 (B); 9.5 ml A is made to 1 l with B, phosphate buffer pH 5.0; phosphate buffer— ethanol (80:20), 1 ml/min	UV 230 and 280 nm	0.5 μg/ml	Plasma
Trimethoprim, sulphamethoxazole + metabolite (N-acetyl- sulphamethoxazole)	RP-LC		45% Methanol—water cont. 10 mM phosphoric acid, 2 ml/min	UV 270 nm		Plasma, serum, urine
Urfadyn, see Nitrofurantoin	HPLC					Serum

TABLE 4
ANTICHOLINERGICS AND CHOLINERGICS
See also Antiparkinsonics (Table 12).

Drugs separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Afloqualone	TLC	Silica gel	Benzene—tetrahydrofuran (3:7 or 1:1); tetrahydrofuran—chloroform—acetone—conc. ammonia (15:10:10:1); <i>n</i> -butanol—acetic acid—water (4:1:1)	^{13}C Autoradiography	Urine, plasma, bile	Combination with GC-MS	142, 143	
Aprophen	HPLC	Partisil 5	Methanol—acetone (30:70) with 0.01% triethanolamine, 1 ml/min	UV 254 nm	Serum		144	
Benzonium bromide	GC-MS	3% OV-225, column directly interfaced with the ion source	Methane, 20 ml/min	MS	Below 5 ng/ml	Plasma	Oven 175°C, injector 200°C, source 250°C; separation after oxidation to benzophenone	145
Biperiden, see Antiparkinsonics					Nitrogen-sensitive detection	Plasma, urine	Column 250°C, detector 275°C, injector 300°C	
(—)-1-Cyclopropyl-methyl-4-(3-trifluoromethylthio-5H-dibenzo[<i>a,d</i>]cyclohepten-5-ylidene)piperidine	GC	3% OV-17 on Gas-Chrom Q, 80–100 mesh	Helium, 30 ml/min					
Edrophonium, see Pyridostigmine								
Galanthamine hydrobromide	HPLC	Polygosil 60-C ₁₈ , RP (5 μm)	Dichloromethane— <i>n</i> -hexane—ethanolamine (500:500:0.5), 1 ml/min	UV 235 nm	5 ng/ml	Serum, urine, bile		147
	HPLC	CP TM Micro-Sphær Si (3 μm) Chrompack	Dichloromethane— <i>n</i> -hexane—ethanolamine (500:500:0.5), 1 ml/min	UV 235 nm	5 ng/ml	Serum, urine, bile		147
Neostigmine, see Pyridostigmine	HPLC	LiChrosorb Si 60 (5 μm)	10% 1 M Ammonium acetate pH 10 in methanol, 1 ml/min	UV 254 nm	Below 0.5 ng/ml	Serum, urine		148

Proglumide (xylamide)	HPLC	Zorbax SII	Chloroform-methanol (24:1), UV 240 nm 1.5 ml/min	0.05 µg/ml	Plasma	149
Propantheline	HPLC	µBondapak C ₁₈ (10 µm)	Acetonitrile-0.1 M KH ₂ PO ₄ pH 3.0 (4:6), 1.5 ml/min	UV 210 nm Below 5 ng/ml	Serum	150
Pyridostigmine	HPLC	µBondapak C ₁₈ (10 µm)	37.5% Acetonitrile in water cont. 0.001 M sodium dodecyl sulphate and 1% acetic acid, final pH 4.0, 1 ml/min	UV 269 nm Below 20 ng/ml	Plasma	151
Pyridostigmine, neostigmine, edrophonium + metabolites	HPLC	LiChrosorb RP-18 (10 µm)	0.01 M C ₆ H ₅ SO ₃ ⁻ Na ⁺ and 0.01 M NaH ₂ PO ₄ in acetonitrile-water (15:85), pH 3.0, 2 ml/min	UV 214 nm Below 5 ng/ml	Serum	152
	HPLC	Ultrasphere-octyl	0.01 M C ₆ H ₅ SO ₃ ⁻ Na ⁺ , 0.01 M NaH ₂ PO ₄ and 0.025 M trimethylamine chloride in acetonitrile-water (20:80), pH 3.0, 2 ml/min	UV 214 nm Below 5 ng/ml	Serum	152
	HPLC	Ultrasphere-octyl	Acetonitrile-water (17:83)	UV 214 nm Below 5 ng/ml	Serum	152
Tacrine (tetrahydroaminoacridin)	HPLC	µBondapak C ₁₈ (5 µm), 1.5 ml/min	0.1 M Phosphoric acid pH 2.8-acetonitrile (16:2), 1.5 ml/min	UV 254 nm or fluorescence at 385/425 nm 100 ng/ml	SepPak C ₁₈ used for extraction Plasma	153
Tetrazepam + metabolites	HPLC	RPC ₁₈ column	Acetonitrile-water (40:60) cont. 5 mM perchloric acid and 10 mM sodium perchlorate Toluene—acetone (9:1)	UV 254 nm Urine	Urine	154
	TLC	Silica gel			Separation of metabolites, combination with IR	154

TABLE 5
ANTICOAGULANTS

Drugs separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Acenocoumarol (Sintrom)	HPLC	LiChrosorb RP-2 (10 μ m)	0.75 g Ammonium acetate in 100 ml acetonitrile—water— acetic acid (37:62:1), 40 ml/h	UV 305 nm	Nanogram range	Plasma		155
	HPLC	LiChrosorb RP-8 (5 μ m)	0.1% Acetic acid—acetonitrile— ethyl acetate pH 4.9 (100:90:1), 1.5 ml/min	UV 303 nm	20 ng/ml	Plasma		156
	HPLC	μ Bondapak C ₁₈ (10 μ m)	Ethanol—water (1:1) cont. 1% of 98% acetic acid, 1.4 ml/min	UV 313 nm	10 ng/ml	Plasma		157
Acenocoumarol + reduced derivative (Aminosintrom)	TLC	Silica gel	Ethyl acetate—methanol—tri- ethanolamine (70:30:3)	UV 366 nm	0.01 μ g/ml	Plasma	Layer immersed into fluores- amine solution; reduction of Sintrom by SnCl ₂ to Amino- sintrom	158
	TLC	Silica gel	(A) Light petroleum—acetone (140:60); (B) benzene—ethyl acetate (140:60)	UV 254 nm; then elution and liquid scintillation		Plasma, urine	¹⁴ C	159
Bisoumacetate	HPLC	μ Bondapak C ₁₈	Methanol—water—acetic acid (56:40:4), 1 ml/min	UV 254 and 280 nm	Below 0.1 μ g/ml	Plasma		160
Brodifacoum, see Warfarin								
Difenacoum, see Warfarin								
Diphenadione (diphasicnone)	TLC	Silica gel	(A) Toluene—methanol—diethyl UV and auto- amine (70:20:10); (B) chloro- form—methanol—water (80:20:4); (C) toluene— acetone—acetic acid (75:20:5); (D) toluene—dioxane—acetic acid (76:21:3)			Urine, faeces, liver	¹⁴ C	161
Ethyl bisoumacetate, see Warfarin								
Nafazatrom	HPLC	LiChrosorb Si 60 (10 μ m)	Dichloromethane—methanol (90:10) cont. 0.25% water, 1.0 ml/min	UV and fluorescence at 232/362 nm	5 ng/ml	Plasma		162

Phenindione (Pindione)	GC	10% SE-30 on Chromosorb W NAW HMDS, 60–80 mesh	Nitrogen, 35 ml/min	FID	10 µg/ml	Plasma	Column 270°C, injector 280°C, detector 300°C	163
Phenprocoumon, (see also Warfarin)	HPLC	LiChrosorb RP-18 (10 µm); pre-column Bondapak C ₁₈ Corasil (35–50 µm)	Plasma: acetonitrile—water—acetic acid (600:400:5), 2.0 ml/min; urine: gradient elution acetonitrile—water—acetic acid (400:600:5) (A); acetonitrile—acetic acid (1000:5) (B); linear gradient from 0 to 1.00% B in 30 min, 2.0 ml/min	0.1 µg/ml (plasma); 0.02 µg/ml (urine)	Plasma, urine			164
TLC	Silica gel		Methanol—triethylamine (80:20) for cleaning the plate; then chloroform—methanol—triethylamine (95:15:6)	UV 254 nm	5 µg/ml	Plasma	Lover edge of the plate is impregnated with pentane—triethylamine (100:10) to stabilize phenprocoumon	165
Prevican (Fluorindione)	GC	10% SE-30 on Chromosorb W NAW HMDS, 60–80 mesh	Nitrogen, 35 ml/min	FID	10 µg/ml	Plasma	Column 270°C, injector 280°C, detector 300°C	163
Warfarin, phenprocoumon	GC	(A) 3% OV-1; (B) 3% OV-17; (C) 3% SP-1000; all on Chromosorb W HP, 100–120 mesh	Nitrogen, 20 ml/min	FID	1 µg/ml	Plasma	Column for warfarin: (A and B) 250°C, (C) 270°C; for phenprocoumon: (A) 230°C, (B) 250°C, (C) 270°C; detector and injector 30°C higher	64
	GC	3.8% Vinyl methyl silicone on Gas-Chrom W HP, 80–100 mesh	Helium, 60 ml/min	FID	0.3 µg/ml	Plasma		166
Warfarin, warfarin alcohol	GC-MS			SIM		Plasma		167
Warfarin, phenprocoumon	HPLC	µBondapak C ₁₈ (10 µm)	Water cont. 0.1% of 99.8% acetic acid—ethanol (1:1), 1.5 ml/min	UV 254 nm	MS	Plasma		168
Warfarin	HPLC	Partisil (5 µm)	n-Hexane—ethanol (93.5:6.5), 1.5 ml/min	UV 280 nm	Below 0.05 µg/ml	Serum, plasma		169
							(Continued on p. 426)	170

TABLE 5 (continued)

Drugs separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Warfarin, difenacoum, brodifacoum, (<i>cis</i> -, <i>trans</i> -isomers)	HPLC	Magnumil 8H C ₁₈ , RP (5 μm)	Methanol, 1 ml/min; cyclohexane—dichloromethane—acetic acid (75:25:0.6), 1 ml/min; or RP chromatography: methanol—water—acetic acid (80:20:0.8), 1 ml/min	Fluorescence at 315/410 nm, UV 260 nm	0.05—0.1 μg/g warfarin; 0.02 μg/g other compounds	Liver, stomach, serum, urine	If analysis of warfarin is not required, sensitivity can be about 1 pg/g	171
Warfarin	HPLC	μBondapak C ₁₈	Methanol—acetic acid—water (75:0.5:25), 1 ml/min	UV 313 nm	Below 0.3 μg/ml	Plasma		172
Warfarin + metabolites (diastereomeric warfarin alcohols, 4-, 6-, 7-, 8-hydroxywarfarin)	HPLC	Partisil-10 PAC (10 μm)	75% <i>n</i> -Heptane—25% solution cont. 79.3% 1,2-dichloroethane, 20% ethanol and 0.7% acetic acid, 2 ml/min	Fluorescence at 318/393 nm (295 nm shoulder)	0.18 ng	Urine, plasma	Post-column reagent solution: 10% triethylamine and 90% solution cont. 75% <i>n</i> -heptane, 20% 1,2-dichloroethane and 5% ethanol	173
Warfarin, acenocoumarol, ethylbisoumaracetate	HPLC	Silanized LiChrosorb Si 60	0.2 M Na ₂ HPO ₄ —acetonitrile (75:25) cont. 0.01 M tetra-butylammonium bromide			Plasma		174
	HPLC	μBondapak C ₁₈	Ethyl acetate— <i>n</i> -hexane—methanol—acetic acid (25:7:4.75:0.25:0.4), 1 ml/min	UV 313 nm	0.06—9.0 μg/ml	Plasma	Separation of isomers <i>R</i> (+)	175
	HPLC	Spherisorb Si (5 μm)	Methanol—water pH 4.3 (63:37)	UV 254 nm	0.16 μg <i>S</i> (—); 0.036 μg <i>R</i> (+)	Plasma	and <i>S</i> (—); esterification with carbobenzoyloxy-L-proline	176
	HPLC	RP			0.5 μg/ml	Plasma		177

TABLE 6

ANTIDIABETICS

Drugs separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Acetohexamide, hydroxyhexamide (see also Tolbutamide)	LC	LiChrosorb	0.5% Acetic acid-acetonitrile (1:1)			Plasma		178
Carbutamide (see also Tolbutamide)	HPLC	Styrene-divinyl- benzene copolymer, ammonium-sub- stituted (Hitachi 3011-N)	0.1 M Sodium hydroxide-0.1 M sodium chloride in 65% methanol, 1.1 ml/min	UV 254 nm		Serum		179
Chlorpropamide, see Glipizide, Tolbutamide								
Cigitazone + monohydroxy metabolites	HPLC	Supelcoil LC-18 (5 μm); pre-column Sphere-5 RP 18	Acetonitrile-7 mM phosphoric acid pH 2.5 (4:6 or 7:3)	UV 229 nm	0.05 μg/ml	Serum		180
Glibenclamide, see Glyburide	HPLC	LiChrosorb C ₁₈ RP (10 μm) 50°C	Methanol-water (7:3)	UV 230 nm	0.5 μg/ml	Serum		181
Glibornuride (see also Glyburide, Tolbutamide)	GC	3% XE 60 on Chromosorb W AW DMCS, 80-100 mesh Silanized glass column; 2% OV-101 on Chromosorb, W AW DMCS, 100-200 mesh	Nitrogen, 30 ml/min Helium, 30 ml/min	⁶⁵ Ni ECD	0.02 μg per sample	Plasma	Derivatization with diazo- methane and heptafluorobutyric anhydride; column 220°C, injector and detector 280°C; Column 220°C, injector 250°C, detector 300°C	182
Gliclazide	GC			NPD				
Glipizide	HPLC	Jasco RP SC-QL (7 μm)	Methanol-0.2% acetic acid (3:2) Acetonitrile-methanol-1.2 M ammonium perchlorate (4:3:7), 0.4 ml/min	ECD UV 227 nm UV 227 nm	0.2 μg/ml 0.2 μg/ml	Plasma Plasma		183
Glipizide	HPLC	Daison CDR-10 Sc-01 and CDR-10 TLC Silica gel	Chloroform-acetone-acetic acid (13:6:1); ethyl acetate- chloroform-acetic acid (3:2:1)	Autordiography	0.2-0.3 μg/ml	Serum Plasma	¹ H	184
								185
								186

(Continued on p. 428)

TABLE 6 (continued)

Drugs separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Glibipizide (Glibenclamide), chlorothiobutamide, chlorpropamide, glyburide (see also Tolbutamide)	HPLC	μ Bondapak C ₁₈ (10 μ m)	Methanol-0.01 M phosphate buffer pH 3.5 (60:40)	UV 225 nm (glibipizide)	10 ng/ml glipizide, other drugs approx. several μ g/ml	Serum	Only benzene is suitable for extraction; 3-cis-hydroxy-cyclohexyl and 4-trans-hydroxy-cyclohexyl metabolites interfere	187
Glibipizide + metformine and sulfonylurea splitting products	TLC	Silica gel	Cyclohexane-ethyl acetate-acetic acid (60:40:2); chloroform-methanol-acetic acid (100:3:1); cyclohexane-ethyl acetate (70:30)	Fluorescence at 336/510 nm	Below 10 ng per spot	Serum	Dns derivatives	188
Glyburide (glibenclamide, gliburone, Maninil), tolbutamide 80-100 mesh	GC	5% OV-17 on Gas-Chrom G AW DMCS, (5 μ m)	Argon-methane (95:5), 100 ml/min	*NI ECD	Below 5 ng/ml	Plasma		189
Glyburide (glibenclamide, gliburone, Maninil), tolbutamide 80-100 mesh	HPLC	LiChrosorb RP-18 (5 μ m)	Acetonitrile-12 mM perchloric acid (47:53), 2.0 ml/min (A); acetonitrile-5 mM phosphoric acid, 2 ml/min (B); acetonitrile-phosphate buffer pH 6, 2 ml/min (B); acetonitrile-perchloric acid, 2 ml/min (B)	UV 230 nm	10 ng/ml	Plasma	(A) Separation of contaminants from serum; after 9 min programming to 100% acrylonitrile during 2 min; (B) determination of the drugs	190
Glyburide, chlorpropamide, chlorothiobutamide, chlorpropamide, glibenclamide, gliburide, Maninil, tolbutamide	HPLC	LiChrosorb Si-60 (10 μ m)	99% Ethanol-n-heptane-diisopropyl ether (30:60:400), 2 ml/min	Fluorescence at 438/522 nm	5 ng/ml	Serum	Pre-column derivatization with NBD-chloride (7-chloro-4-nitrobenzofuran)	191
Glyburide, chlorpropamide, chlorothiobutamide, chlorpropamide, glibenclamide, gliburide, Maninil, tolbutamide	HPLC	LiChrosorb RP-18 (5 μ m)	Acetonitrile-0.05% phosphoric acid (45:55), 1.1 ml/min	UV 230 nm	Below 50 ng/ml	Serum		192
Glyburide, chlorpropamide, chlorothiobutamide, chlorpropamide, glibenclamide, gliburide, Maninil, tolbutamide	HPLC	Spherisorb ODS (5 μ m)	Acetonitrile-1/15 M phosphate buffer pH 7 (2:5), 2 ml/min	UV 228 nm	5 ng/ml	Serum		193
Glyburide, chlorpropamide, chlorothiobutamide, chlorpropamide, glibenclamide, gliburide, Maninil, tolbutamide	HPLC	LiChrosorb RP-8 (5 μ m)	buffer pH 7 (2:5), 2 ml/min	UV 228 nm	20 ng/ml	Serum		194
Glyburide, chlorpropamide, chlorothiobutamide, chlorpropamide, glibenclamide, gliburide, Maninil, tolbutamide	HPLC	LiChrosorb RP-8 (10 μ m)	(1:1), 1.2 ml/min	Fluorescence at 380/360 nm	10 ng/ml	Serum		195
Glyburide, chlorpropamide, chlorothiobutamide, chlorpropamide, glibenclamide, gliburide, Maninil, tolbutamide	HPTLC	Silica gel	Chloroform-methanol-conc. ammonia (15:3:0.2)	Reflectance mode scanning at 300 nm	5 ng/ml	Serum		196
Glyburide, chlorpropamide, chlorothiobutamide, chlorpropamide, glibenclamide, gliburide, Maninil, tolbutamide	HPLC	Magnusphere C ₁₈ , RP (5 μ m)	0.05 M Ammonium phosphate-methanol (38:62), 0.8 ml/min	UV 225 nm	2 ng/ml	Serum		197

[3] **Woodiazine** see Tollutamide

Metformin	GC	3% OV-17 on silanized Chromosorb W, 80–100 mesh	Nitrogen, 40 ml/min	Nitrogen-sensitive detection (KCl salt)	25 ng/ml	Plasma	Oven 250°C; derivatization with <i>p</i> -nitrobenzoyl chloride	199
Phenformin (Dipar)	HPLC	μ Bondapak C ₁₈	Methanol—water (1:1) cont. 0.02% acetic acid and 5 · 10 ⁻³ M 1-heptanesulfonic acid, 1 ml/min	UV 235 nm	10 ng/ml (plasma); 250 ng/ml (urine)	Plasma, urine		200
Tolazamide, see Tolbutamide								
Tolbutamide, chloropropanide, glibizide, glibenclamide, glibornuride, tolazamide	GC	3% OV-17 on Gas-Chrom Q, 100–120 mesh	Nitrogen, 40 ml/min	FID, ECD	20 ng/ml	Plasma	Methyl derivatives; injector and detector 330°C, oven 300°C for glipizide derivatives, 220°C for tolbutamide derivatives	201
Tolbutamide, chlorpropamide, carbutamide, tolazamide, glycodiazine	GC	(A) 3% SE-30; (B) 5% SE-30; (C) 2% OV-17; all on Chromosorb W AW DMCS, 80–100 mesh	Helium, 40 ml/min	¹⁴ N ECD	250 ng/ml tolbutamide and chloropropane; 3000 ng/ml carbutamide; 500 ng/ml tolazamide; 1500 ng/ml glycodiazine	Blood	Derivatization with diazo-methane and trifluoroacetic anhydride; column temperatures: (A) 1 min at 195°C, 10°C/min to 250°C, 5 min at 250°C; (B) 1 min at 220°C, 10°C/min to 280°C, 5 min at 280°C; (C) 1 min at 210°C, 10°C/min to 250°C, 5 min at 250°C	202
Tolbutamide, chlorpropamide	HPLC	μ Bondapak C ₁₈ , RP	Acetic acid—acetonitrile (72:28), 2.2 ml/min	UV 254 nm	6 μ g/ml tolbutamide; 7 μ g/ml chloropropane	Serum		203
Tolbutamide + metabolite (carboxytolbutamide)	HPLC	μ Bondapak C ₁₈ , 28°C	Acetonitrile—phosphate buffer pH 3.9 (35:65)	UV 254 nm	2 μ g/ml tolbutamide; 0.1 μ g/ml carboxy-tolbutamide	Plasma		204
Tolbutamide + metabolites (carboxytolbutamide, hydroxymethyl derivative)	HPLC	ODS-Si-X-1 μ Bondapak HPLC	Methanol—0.2% acetic acid (3:2), 1.2 ml/min	UV 228 nm	200 ng/ml	Plasma		200
	HPTLC	Silica gel	Water—acetonitrile (78:22) Acetonitrile—0.05% phosphoric acid (45:55), 1.5 ml/min	UV 200 nm	0.5 μ g/ml	Plasma	100- μ l Aliquots	205 206
	HPTLC		Chloroform—acetic acid—methanol (18:2:1:0.8)	Fluorescence at 366 nm		Plasma		102

TABLE 7
ANTIEMETICS

Drugs separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Alizapride	HPLC	μ Bondapak C ₁₈	Methanol—pH 8.1 buffer (80:20, 2 ml/min)	Fluorescence at 323/380 nm	5 ng/ml	Plasma, urine		207
Bromide (Vabien) + metabolites	TLC	Silica gel	Chloroform—methanol—ammonia (35:20:5 or 35:25:5)			Urine	Isolation of metabolites for GC and MS	208
Butaperazine, trifluopromazine	LC	Micropak CN (1.0 μ m)	90% Acetonitrile or methanol and 1.0% aq. ammonium acetate; its concentration varied from 0.005 M to 0.2 M, 2.0—2.5 ml/min	UV and electrochemical detection	10 ng/ml	Blood, plasma		209
Clebopride, see Metoclopramide								
Cyclizine, norgyzine	GC	5% OV-17 on Chromosorb W HP, 100—120 mesh	Helium, 50 ml/min	NPD	20 pg Cyclizine and 2 ng nor-cyclizine injected on-column	Plasma, urine	Manifold 321°C, oven 246°C, injector 310°C	210
Domperidone + metabolites	HPLC	Lichrosorb RP 8 (5 μ m) or Li-Chrosorb NH ₂ (5 μ m)	Linear gradient 100% 0.1 M ammonium acetate to 100% mixture of 1 M ammonium acetate—methanol—acetonitrile (10:4:5:45) (A) over 30 min, 1 ml/min; or linear gradient 99% diisopropyl ether and 1% A to 40% of this solvent and 60% A, 2.0 ml/min	UV 240 or 290 nm or ¹⁴ C liquid scintillation counting		Plasma, urine, bile		211
Hopantemic acid (homopanthothenic acid)	GC-MS	3% OV-17 on Chromosorb W AW, 80—100 mesh	Helium, 30 ml/min	Ion source	5 ng/ml	Plasma	Column 220°C, injector 250°C, ion source 230°C; when the drug and internal standard had been detected, the temperature of the column raised to 280°C	212
	GC	3% OV-17 on Chromosorb W AW, 80—100 mesh	Nitrogen, 60 ml/min	FID	5 ng/ml		Column 170°C, 6°C/min to 290°C, injector and detector 280°C	

Medazine	GC-MS				Below 5 ng/ml	Plasma	213
Metoclopramide	GC	3% OV-101 on acid-washed DMCs-treated Gas-Chrom Q, 80–100 mesh Fused-silica capillary column, 5% phenylmethylsilicone Ultra No. 2, siloxane-deactivated	Nitrogen, 37 ml/min	Scandium tritide ECD	5 ng/ml	Plasma	Derivatization with heptafluorobutyrylimidazole; column 200°C, injector and foil 275°C
	GC-MS	3% OV-17 on Chromosorb W, 80–100 mesh	Hydrogen, 1.0 ml/min	ECD	Below 4 ng/ml	Plasma	Column 235°C, injector 220°C, detector 350°C
	GC	3% OV-17 on Chromosorb W, 80–100 mesh	Argon—methane (95:5), 40 ml/min	^{63}Ni ECD	1 pg per injection	Plasma, blood, urine	Derivatization with heptafluorobutyric anhydride; oven and injector 250°C, detector 350°C
	HPLC	3% OV-17 on Chromosorb W, 80–100 mesh Nucleosil C ₁₈ (5 μm) 50°C	Helium, 40 ml/min	MS		Plasma, urine	Oven and injector 250°C, separator 200°C, analyzer 50°C
	HPLC	RP-18	Argon—methane (95:5), 40 ml/min	^{63}Ni ECD	7 ng/ml	Plasma, urine	Open 245°C (plasma), 230°C (urine), injector 250°C, detector 350°C
	HPLC	Spherisorb	32% Acetic acid (1% solution)—63% acetonitrile—methanol (3:7:1)	UV 273 nm	8 ng/ml	Plasma	Open 245°C (plasma), 230°C (urine), injector 250°C, detector 350°C
	HPLC	Silica gel M131	40% <i>n</i> -Propanol, 50 mM ammonium nitrate pH 7.6, 1.2 ml/min	UV 308 nm	Below 10 ng/ml	Serum	219
	HPLC	TLC	Dichloromethane—methanol—ammonia (90:10:0.5), 1.7 ml/min	UV 308 nm	5 ng/ml	Plasma, blood, urine	220
			Methanol—chloroform—conc. ammonia (30:70:0.5), 2 ml/min	UV 280 nm	10 ng/ml (plasma or blood)	Plasma, blood, urine	221
			2 M ammonia—ethanol—ethyl acetate (3:3:8); ethyl acetate—acetic acid—water—ethanol (25:12:8:5); chloroform—methanol—conc. ammonia (70:30:1, 70:30:2 or 90:10:1); 1-butanol—ethyl acetate—acetic acid—water (1:1:1:1)				221

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TABLE 7 (continued)

Drugs separated	Method	Stationary phase	Mobile phases or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Metoclopramide, clobopride + metabolites	TLC	Silica gel	Methanol—chloroform (1:4); 1,2-dichloroethane—ethanol— ammonia (sp. gr. 0.88) (70:15:2); <i>n</i> -butanol—acetic acid—water (4:1:1); 2- propanol—ammonia (sp. gr. 0.88)—water (80:40:5)	Photodensito- metry after diazo- coupling with N- (1-naphthyl)- ethylene di- ammonium dichloride	20 ng/ml	Liver and other bio- logical material		222
	HPTLC	Silica gel	Chloroform; development in NH_3 atmosphere	Spectrophoto- metry at 308 nm	36 ng/ml	Serum		223
	HPTLC	Silica gel	Chloroform—acetic acid— methanol (18:2:1:0:0.8)	Fluorescence at 366 nm		Plasma		102
Salipride + N-methyl-, N-propyl derivatives, six metabolites	LC—MS			Direct-probe chemical ionization	Nanogram level	Hypophysis		224
	HPLC	C_{18}	(A) Water—acetic acid (99:1); (B) acetonitrile—acetic acid— water (50:1:49); linear gradient from 12 to 60% B in 10 min, 1.2—1.6 ml/min	Fluorescence at 259/342 nm	10 ng/ml	Serum, urine, CSF		225
Salipride + metabolites	TLC	Silica gel	Chloroform—methanol—25% ammonium hydroxide (60:40:2); 2-propanol— toluene—25% ammonium hydroxide (60:30:10); benzene—methanol—acetic acid (60:35:5); methanol; <i>n</i> -butanol—methanol—water— acetic acid (60:25:25:6)	^{14}C Radioscanning		Faeces		226
	PC	S+S 2043B paper	<i>n</i> -Butanol—water—25% ammonium hydroxide (80:10:10)					226
Triflupromazine, see Butaperazine								
Trimethobenzamide	GC—MS	3% SE on Supelcoport, 100—120 mesh	Helium, 30 ml/min	0.1 $\mu\text{l}/\text{ml}$	Saliva	Column and ion source 250°C, injector 260°C, separator 270°C		227

TABLE 8
ANTIMYCOTICS (OTHER THAN ANTIBIOTICS)

Drugs separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Benzoic acid	GC	3% OV-1 (A); 3% OV-17 (B); 3% SP-1000 (C); all on Chromosorb W HP 100–120 mesh	Nitrogen, 20 ml/min	FID	1 µg/ml	Plasma	Preparation of butyl esters; column temperatures: (A) 120°C, (B) 140°C, (C) 150°C, injector and detector 30°C higher	64
Econazole	HPLC	Partisil 10 ODS RP	Methanol—ac. 0.01 M KH ₂ PO ₄ , pH 4.5 (70:30), 2 ml/min	UV 220 nm	0.04 µg/ml	Plasma		228
Fluconazole (5-fluorocytosine)	HPLC	µBondapak C ₁₈	10 mmol KH ₂ PO ₄ , buffer pH 7.0, 1.5 ml/min	UV 276 nm	Below 5 µg/ml	Plasma, CSF	5-Fluorouracil interferes	229
	HPLC	LiChrosorb RP (18.5 µm); guard column Bondapak C ₁₈ Corasil (37–50 µm)	15 mM Octane sulphonate in acetic acid—acetone—water (5:7:88), 1.1 ml/min	UV 280 nm	Below 5 µg/ml	Serum		230
Ketoconazole	HPLC	µBondapak CN RP	0.05 M KH ₂ PO ₄ —sodium hydroxide buffer pH 6.0—acetonitrile (65:35), 2.0 ml/min	UV 205 nm	0.1 µg/ml	Plasma		231
	HPLC	µBondapak C ₁₈ RP (10 µm)	Acetonitrile—Sorensen's phosphate buffer pH 6.6 (60:40), 1.5 ml/min	Fluorescence at 206/370 nm		Plasma		232
	HPLC	Ultrasphere ODS (5 µm); pre-column Ultrasphere ODS (20 µm)	Water—methanol—diethyl amine (25:75:0.1), 1 ml/min	UV 240 nm	ca. 0.1 µmol/l	Plasma, serum		233
	HPLC	Altex Ultrasphere octadecylsilane; pre-column Whatman C ₁₈ (30—36 µm)	Methanol—0.02 M monohaiic sodium phosphate (75:25), 1.0 ml/min	UV 231 nm	0.2 µg/ml	Serum		234
Miconazole	HPLC	µBondapak C ₁₈ RP 50	Methanol—tetrahydrofuran—2.5 mmol/l acq. acetate buffer pH 5 (0.25:1:32.5), 2.0 ml/min	UV 254 nm or 280 nm		Plasma	Mobile phase contained 6 mmol/l sulphonate	235
	HPLC	RadialPak C ₁₈ ; pre-column Bondapak C ₁₈ Corasil	77% Methanol in 0.01 M EDTA with 0.005 M n-nonylamine, 1.5 ml/min	UV 230 nm	0.5 µg/ml	Saliva		236

(Continued on p. 434)

TABLE 8 (continued)

Drugs separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Oxiconazole	GC	3% SP-2250 on Supelcoport, 80-100 mesh	Nitrogen, 40 ml/min	^{63}Ni 10 mCi ECD or ^{63}Ni 15 mCi ECD	1 ng/ml	Plasma	Column 270-290°C, injector 300°C, detector 350°C	237
Salicylic acid; see Antibacterials								
Sulconazole	HPLC	μ -Bondapak C ₁₈ RP; pre-column Co ₂ Pell ODS Whatman	Acetonitrile-0.01 M NaH ₂ PO ₄ , buffer pH 8.0 (66:34), 2 ml/min	UV 229 nm	Less than 0.5 µg/ml	Plasma		238

TABLE 9
ANTIHISTAMINES

See also Antiallergic drugs (Table 14). Cross-references between individual types of antihistamines are not supplied. Check the whole table for completeness.

Drug separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Astemizole	HPLC	R Sil C., HL (5 μm)	Acetonitrile—water (50:50), 0.6 ml/min	UV 254 nm	1 ng/ml (plasma); 5 ng/ml (other biological material)	Plasma, tissue		289
Azelastine	GC-MS	Slanized glass column, 3% OV-1 on Shimalite W, 80–100 mesh	Helium, 20 ml/min	Multiplication detection	0.5 ng/ml	Plasma	Column 250°C, flash heater 310°C, ion source 330°C	240
Bromodiphenylhydramine (Promazine) + metabolites	TLC	Silica gel	Benzene; chloroform—acetone (90:10); acetone—25% ammonia (100:1); chloroform—methanol—ammonia (90:10:1); benzene—methanol (98:4); chloroform—acetone (98:2)	Chemical detection		Urine	Isolation and identification of metabolites	241
Carboxinamine	GC	Capillary fused-silica open column, coated with SE-30	Helium	Nitrogen-sensitive detection	0.2 ng/ml	Serum	Oven programmed 185°C to 250°C (10°C/min), detector 300°C, injector 260°C	242
Chlorpheniramine	GC	3% OV-17 on Chromosorb Q AW DMCS, 100–120 mesh	Nitrogen, 1.68 kg/cm ²	FID	Below 60 ng	Urine	Oven 210°C, injector 260°C; column silanized	243
		2% OV-101 on Chromosorb W HP, 120 mesh	Helium, 30 ml/min	NPD	1 ng/ml	Plasma	Column 215°C, injector 200°C, detector 300°C	244
Chlorpheniramine + metabolites	GC	3% SP-2250 phenyl-methylsilicone oil on Supelcoport, 100–120 mesh; also studied 2% OV-101 on Chromosorb W, 100–120 mesh; 3% OV-17 on Gas-Chrom Q, 80–100	Helium, 20 ml/min	NPD	0.5 ng/ml	Plasma	Column 210°C, injector and detector 300°C	245

(Continued on p. 436)

TABLE 9 (continued)

Drugs separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Chlorphenamine	GC	mesh; 3% Carbowax 20 M (pretreated with 5% KOH); 80–100 mesh	Nitrogen, 35 ml/min	FID, nitrogen- sensitive detection	Below 2.5 µg	Urine	Simultaneous determination with phenylpropanolamine (sympathomimetics); injector 240°C, detector 280°C, column 230°C	246
	HPLC	µBondapak C ₁₈ (10 µm)	For plasma and saliva: acetone- nitrile—phosphate buffer 0.075 M NH ₄ H ₂ PO ₄ in 0.16% phosphoric acid (20:80); for urine: 0.05 M NH ₄ H ₂ PO ₄ in 0.11% phosphoric acid; both 2 ml/min	UV 254 nm	Below 2 ng/ml	Plasma, urine, saliva		247
	HPLC	µBondapak C ₁₈ RP < 10 µm)	20% Acetonitrile in phosphate buffer (0.075 M monobasic ammonium phosphate in 0.016 M phosphoric acid, 1 ml/min)	UV 264 nm	UV 264 nm	Plasma	Influence of docosanoate sodium on the release of chlor- phenamine	248
	HPLC	µBondapak C ₁₈ RP (10 µm)	A. Acetonitrile—0.075 M mono- basic ammonium phosphate pH 2.6 (1:4) 1.0 ml/min B. Acetonitrile in 0.075 M phosphate buffer, 2 ml/min	UV 254 nm	1 ng/ml	Plasma		249
	HPLC	µBondapak C ₁₈ RP				Urine		250
Chlorphenamine + metabolites	TLC	Silica gel	Ethyl acetate—methanol—conc. ammonia (10:9:1); benzene— diethyl ether—glacial acetic acid—methanol (60:30:9:1); chloroform—acetone (7:1); methanol; ethyl acetate— methanol (1:1); chloro- form—acetone (3:1); toluene—acetone (3:2)	UV, fluorescamine, ¹⁴ C autoradiog- raphy	UV, fluorescamine, ¹⁴ C autoradiog- raphy	Urine	Methylation of polar metabolites	251
Chlorpromazine, haloperidol	HPLC	MicroPak CN (10 µm)	10% 0.005 M Ammonium acetate in methanol	OD 254 nm	1 ng/ml	Plasma		252
Chlorpromazine	HPLC	LiChrosorb C ₁₈ RP (10 µm); pre- column Co-Pell ODS (30–38 µm)	4.2% Acetonitrile and 3% n- nonylamine in 0.02 M phosphate buffer pH 2.5, 2 ml/min	UV 254 nm	1 ng/ml	Plasma		253

Cinnarizine	HPLC	Spherisorb 5 ODS	Methanol—aq. 0.05 M ammonium dihydrogen phosphate (550:250), 1 ml/min	UV 285 nm	2 ng/ml	Plasma	254
		LiChrosorb RP-8 (5 μ m); pre-column LiChrosorb RP-20 (30 μ m)	Methanol—sodium acetate buffer pH 5.2 (85:15), 2.0 ml/min	UV 250 nm	2 ng/ml	Plasma	255
Cyproheptadine	GC	3% SP-2250 on Supelcort, 80–100 mesh μ Bondapak (10 μ m), 40°C	Helium, 30 ml/min 37% Acetonitrile in 0.1% phosphoric acid, 2 ml/min	Nitrogen-sensitive detector UV 285 nm	Below 3 ng/ml	Plasma, urine	Column 230°C, detector 300°C, injector 275°C
	HPLC	10% Apiezon L-2%	Nitrogen, 30 ml/min	FID	About 4 ng/ml	Urine, plasma	257
Dimethindene	GC	KOH on Chromosorb W AW, 80–100 mesh	Helium, 30 ml/min	Nitrogen-sensitive detector	Below 0.2 μ g/ml	Serum, urine	Column 240°C, injector 200°C, detector 250°C
Diphenhydramine (see also Antiparkinsonics, Orphenadrin)	GC	3% SP-2250 on Supelcort, 100–120 mesh	Helium, 30 ml/min	NPD	1 ng/ml	Serum	Injector 250°C, detector 300°C, column programmed 130–250°C (8°C/min); drugs were absorbed from 1 ml of serum onto XAD-2 resin
Diphenhydramine	GC	3% SP-2250 on Supelcort, 80–100 mesh	Helium, 30 ml/min	NPD	1 ng/ml	Plasma	Injector 310°C, column 205°C, detector 275°C
Doxepin	GC	3% SP-2250 on Supelcort, 100–120 mesh	Helium, 30 ml/min	Nitrogen-sensitive detector	Below 0.2 μ g/ml	Serum	Injector 250°C, detector 300°C, column programmed 130–260°C (8°C/min)
Doxylamine	HPLC	μ Porsil (10 μ m)	8 Parts chloroform, 1 part acetonitrile and 1 part mixture methanol—ammonium hydroxide—ammonium chloride (57:2:1), 1.5 ml/min	UV 254 nm	Below 5 ng/ml	Plasma	261

Mecazine, see Antiemetics

(Continued on p. 438)

TABLE 9 (continued)

Drugs separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Methapyrilene (see also Pyrilamine)	GC	2% OV-1 on Chromosorb G HP, 100–120 mesh	Nitrogen (extra dry), 60 ml/min	NPD	Below 2 ng/ml	Plasma,	Oven 230°C, injector 250°C, detector 300°C	262
	GC	3% SP-2250 on Supelcort, 100–120 mesh	Helium, 30 ml/min	Nitrogen-sensitive detection	Below 0.2 µg/ml	Serum	Injector 250°C, detector 300°C, column programmed 130–260°C (8°C/min)	259
Metiamide	TLC	Silica gel	Ethyl acetate—methanol— ammonia (5:1:1 or 10:1:1); propanol—ammonia (50:1)	UV 254 nm, ³ H scintillation		Urine		263
2-Methoxy-11-oxo- 11H-pyrido[2,1- <i>b</i>]- quinoxoline-8-car- boxylic acid	HPLC	µBondapak C ₁₈ , RP (10 µm)	2 Ampoules PIC reagent A, each cont. 0.0005 M tetraethyl- ammonium phosphate in 14 ml phosphate buffer pH 7.5 per 1 l methanol—water (1:1), 2.0 ml/min	UV 265 nm, fluorescence at 360/475 nm or 280/418 nm	100 ng/ml (UV); 5 ng/ml (fluorescence)	Blood, plasma, urine		264
trans-3-[6-(Methyl- thio)-4-oxo-4H- quinazolin-1-3- propanoic acid	HPLC	µBondapak C ₁₈ (10 µm)	Acetonitrile—methanol— 0.001 M ascorbic acid pH 3.25 (36:28:36), 1 ml/min	Fluorescence at 245/418 nm	0.125 µg/ml	Plasma		265
Mianserin	GC-MS	1% UXR on Gas-Chrom Q	Helium	MS		Plasma	Column 210°C, injector and detector 270°C	266
	GC	3% SP-2260 on Supelcort, 80–100 mesh	Nitrogen, 20 ml/min	NPD	1 ng/ml	Plasma	Oven 230°C, injector and detector 260°C	267
Mianserin + metabolites	LC	Partisil-10 ODS-3 (10 µm) or LC-1, Supelco (5 µm)	0.1 M Acetate buffer pH 4.2— acetonitrile (67:37) with 0.005 M sodium heptane sulphonate, 1.5 ml/min	Electrochemical detector	5 ng/ml	Plasma	Chlormazoline, its metab- olites and hydroxylated metabolites of loxapine interfere	268
Oxaprotiline R(–) and S(+)	HPLC	LiChrosorb Si 60 (10 µm)	1,2-Dichloroethane— <i>n</i> - heptane—ethanol (82:15:3), 4 ml/min	UV 260 nm, ¹⁴ C liquid scintillation counting	2 ng/ml	Blood, urine	Enantiomers were reacted with optically pure N-trifluoroacetyl- <i>S</i> (\leftarrow)prolyl chloride	269
Oxmetidine	HPLC	(A) Ultrasphere Si (5 µm); (B) Ultra- sphere ODS RP (5 µm) 75°C	(A) Acetonitrile—methanol— water—ammonium hydroxide (sp. gr. 0.88) (200:40:10:1.5), 1.8 ml/min; (B) acetonitrile— 0.02 M camphorsulphonic acid (30:70), 2 ml/min	UV 226 nm	Below 5 ng/ml	Plasma, urine, bile		270

Ormetidine + sulfoxide metabolite	HPLC	Ultraphere ODS (5 μ m)	Water-methanol-acetonitrile (45:44:11) cont. 0.095 mol/l pentanesulphonic acid, 1.5 ml/min	UV 226 nm	0.5 μ g/ml (oxmetidine), below 0.25 μ g/ml (metabolite)	Plasma, urine, bile	271	
Promethazine	HPLC	LiChrosorb Si 60 (5 μ m)	Acetonitrile-water-ammonium hydroxide (2500:125:1.5)	UV 228 nm	0.01-0.02 μ g/ml	Plasma, urine	272	
Promethazine + metabolites	GC-MS	3% Carbowax K 20M + 1% KOH on Chromosorb W AW DMCS, 80-100 mesh	Nitrogen, 12 ml/min	Nitrogen-sensitive detection, MS	5 μ g/g (ml)	Liver or kidney homogenate, blood, plasma, urine	Open 220°C, injector 270°C, detector 300°C	273
Promethazine + metabolites	GC	3% OV-17 on acid-washed Chromosorb W, 80-100 mesh	Nitrogen, 4.0 ml/min	Rubidium bead alkaline FID	Alt-glass silanized; column 260°C, injector and detector 270°C	Plasma, urine	274	
Promethazine, trimipazine, 21 compounds of phenothiazine, thioxanthine, dibenzazepine- and butyropheno-like structures	HPLC	Hyperil 5 SAS	Methanol containing 3.0% 0.05 M Sorensen's phosphate buffer pH 7.4, 0.7 ml/min	UV 24.8 nm	0.2 ng/ml	Blood	275	
Promethazine, 12 compounds of phenothiazine-, thioxanthine- and butyropheno-like structures	LC	MicroPak CN (10 μ m)	90% Acetonitrile or methanol and 10% ac. ammonium acetate (conc. varied from 0.005 M to 0.2 M), 2.0-2.5 ml/min	UV and electrochemical detection	10 ng/ml	Blood, plasma	209	
Promethazine	HPLC	MicroPak CN	10% 0.005-0.1 M Ammonium acetate in methanol or acetonitrile	Electrochemical detection		Plasma	276	
Promethazine	LC	MCH-10 RP Varian	Methanol-water (84:16), 2 ml/min	UV 254 nm	1 ng/ml	Serum	277	
Promethazine	HPLC	LiChrosorb C ₁₈ RP (10 μ m); pre-column Co: Pell ODS (30-38 μ m)	4.2% Acetonitrile and 3% <i>n</i> -nonylamine in 0.02 M phosphate buffer pH 2.5, 2 ml/min	UV 254 nm	ca. 1 ng/ml	Plasma	253	

(Continued on p. 440)

TABLE 9 (continued)

Drug separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.	
Proxicronil + metabolites	HPLC	Hi Chrom 5 SODS HPLC	Stepped gradient; for metabolites: methanol—0.5% ammonium acetate (45:55); for proxicronil: methanol—ammonium acetate (60:40); Chloroform—diethyl ether—formic acid (7:2:1)	UV 260 nm		Urine, faeces		278	
Pyrilamine, triprolidine, methapyrylene	TLC	Silica gel	Helium, 25 ml/min	Rubidium bead NPD	10 ng/g (waste water)	Feed-stuffs, urine, waste water	Injector 220°C, oven 200°C, detector 280°C	279	
	HPLC	Altex (5 µm) Si Ultrasphere (for pyrilamine and methapyrylene); Altex (5 µm) Ultrasphere ODS (for triprolidine)	Dichloromethane—2-propanol (99.5:0.5) cont. 0.005 M triethylamine, 2 ml/min	Fluorescence at 310/360 nm	10 µg/g (feed); 1 ng/g (urine)				
			Methanol—0.01 M monobasic potassium phosphate buffer (90:10) cont. 0.005 M terramethylammonium hydrogen chloride pH 7.0, 1.0 ml/min	UV 254 nm					
Pyrilamine							A review with 263 references including analytical methods	280	
SK 8F 934/79 i.e. 2-[2-[5-(dimethylaminomethyl)furan-2-ylmethylthio]ethylamino]-5-(6-methyl-pyridin-3-ylmethyl)-4-pyrimidin-4-one	HPLC	Ultrasphere Altex (5 µm)	Acetonitrile—methanol—water— ammonia (sp. gr. 0.88), (200:80:10:1.5), 2 ml/min	UV 220 nm	0.025 µg/ml	Plasma		281	
Sodium cromoglycate	Anion-exchange HPLC	Partisil SAX (10 µm)	Phosphate buffer (pH 2.3), 36 ml/min	UV 325 nm	About 0.2 µg/ml	Urine		282	
'Trimeprazine, see Promethazine									
Tripelennamine, pentazocine	GC	3% OV-17 on Chromosorb W HP, 80–100 mesh	Nitrogen, 30 ml/min	Nitrogen-sensitive detector			Blood	Column 220°C, injector 250°C, detector 275°C; interference of chloropheniramine and methapyrylene	283
Triprolidine (see also Pyrilamine)	TLC	Silica gel	Methanol—ammonia—chloroform (10:1:39)	Fluorimetry at 300/405 nm	0.8 ng/ml	Plasma		284	

TABLE 10
ANTIMALARIALS

Drugs separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Amodiaquine, see Chloroquine								
Chloroquine (Proguanil), cyclo- guanil + metabolite (4-chlorophenylbi- guanide)	HPLC	Hyperil ODS (5 μ m)	Acetonitrile—water (1:1), cont. 0.17 M phosphoric acid and 0.0125 M NaH ₂ PO ₄ , 1.5 ml/min	UV 247 nm	60 ng/ml	Serum	Lauryl sulphate as pairing ion	285
Chloroquine, pyrimethamine, quinine, sulphadoxine	GC	2% OV-17 on Chromosorb W AW DMCS, 100–120 mesh	Nitrogen, 30 ml/min	NPD (thermoionic)	0.23 ng/ml chloroquine; 0.52 ng/ml quinine; 0.191 ng/ml pyrimethamine;	Blood, urine	Column programmed 200– 290°C (8°C/min), detector 350°C, injector 250°C	286
	GC	3% OV-1—OV-17 (1:3) on Gas-Chrom Q, 100–120 mesh	Nitrogen, 50 ml/min	Nitrogen-sensitive detection	2.31 ng/ml sulphadoxine 20 ng/ml	Blood	Column 235°C, injector and detector 300°C	287
Chloroquine + metabolite	GC-MS	3% OV-17 on Gas-Chrom Q, 80–100 mesh	Nitrogen, 40 ml/min	MS	0.2 μ mol/l	Plasma, urine	Derivatization with trifluoro- acetic anhydride; column (silanized) 250°C, injector and detector 300°C	288
Chloroquine + metabolite	GC	Fused-silica capillary column, siloxane-deacti- vated and coated with OV-1	Helium, 3–4 ml/min	Nitrogen-sensitive detection	5 ng/ml chloro- quine; 15 ng/ml metabolite	Blood	Perfluorocacylation; column programmed 125–230°C (20°C/min) (method A) or 220°C (method B), injector 125°C, detector 290°C	289
	GC-MS	5% OV-101 on Gas-Chrom Q, 100–120 mesh	Helium, 16 ml/min	MS		Blood	Derivatization with bis(penta- fluoropropionic) anhydride	
	GC-MS	3% OV-17 on Supelcopor, 80–100 mesh	Nitrogen, 30 ml/min	FID, MS	Below 10 μ g/ml	Urine	Column 230°C, injector 240°C detector 265°C, prior to GC TLC on silica in 25% ammonia— methanol (3:200); UV detection	290

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TABLE 10 (continued)

Drugs separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Chloroquine + metabolites	GC	J&W DB-5 and SGE BP-5 fused-silica capillary column	Nitrogen, 1 ml/min	Nitrogen-sensitive detector (thermolonic)	3 ng/ml chloroquine; 10 ng/ml metabolite	Blood	Column 255°C, injector and detector 280°C	291
Chloroquine + metabolite	HPLC	Nucleosil C ₁₈ (5–10 μm) (when drug is injected after extraction from biological material); LiChrosorb RP-8 (10 μm) (when direct injection of biological material is used)	Acetonitrile–phosphate buffer pH 3 (40:60), 0.8–1.0 ml/min	UV 254 or 340 nm; fluorescence at 335 nm (370-nm cut-off filter)	ca. 0.5 nmol/l	Plasma, blood, urine		292
Chloroquine + desethyl metabolite	HPLC	Nucleosil C ₁₈ (5–10 μm); guard column LiChrosorb RP 8	Acetonitrile–phosphate buffer (40:60), 0.8–1.0 ml/min	Fluorescence	0.5 μmol/l	Serum, plasma, blood cells, urine		293
	Ion pair RP-HPLC	Pattis-10 ODS-3 RO	Methanol–water–acetic acid (80:19:1) cont. 0.005 M sodium heptanesulphonate, 1 ml/min	UV 344 nm	20 ng/ml	Blood, urine		294
Chloroquine + desethyl metabolite	LC	LiChrosorb Si 60 (5 μm)	Acetonitrile–methanol–diethylamine (80:19.5:0.5), 1 ml/min	Fluorescence at 335 nm (370-nm cut-off filter)	1 ng/ml chloroquine; 0.5 ng/ml desethyl metabolite	Plasma, red blood cells, urine		295
Chloroquine + desethyl metabolite	HPLC	μBondapak C ₁₈ (10 μm)	0.02 M PIC B-7 reagent–acetonitrile (66:34), 1.0 ml/min	UV 340 nm		Plasma		296
Chloroquine + metabolite	HPLC	Dupont Zorbax Zill (5–6 μm)	Hexane–methyl tert.-butyl ether–methanol–diethylamine (37.25:37.25:0.5), 1.0 ml/min	UV 340 nm, fluorescence at 320/380 nm	5 ng/ml	Blood		297
Chloroquine, amodiaquin, pyrimethamine, quinine	HPLC	Dupont Zorbax Sil (5–6 μm)	Hexane–methyl tert.-butyl ether–methanol–diethylamine (46:46:7.5:0.5)	UV 254 nm		Blood		297

Chloroquine + desethyl- and bis(dieethyl) metabolites	HPLC	LiChrosorb 10 RP 18	Water—acetonitrile (68:32); cont. sodium perchlorate, trimethylammonium chloride and sodium acetate, pH ad- justed to 9.25 with borate buffer, 1 ml/min	UV 200—400 nm Benzene—methanol—diethyl- amine (7.5:1.5:1); chloro- form—ethyl acetate (1:1)	1 ng/ml	Plasma	298	
Cycloguanil, see Chloroguanide	TLC	Silica gel, activated	<i>n</i> -Butanol—acetic acid—water (66:17:17); benzene—methanol (3:1)	Urine, faeces, plasma, blood, tissue samples, expired air			300	
Diminazene	HPLC	Radial Pak CN (10 µm); CN pre-column	Acetonitrile—water (50:50) cont. 0.2% triethylamine pH 4.2	UV 254 nm	About 0.05 µg	Plasma	301	
Mefloquin + DL-threo-α-2'-piperityl- 2-(4-trifluoromethyl- phenyl)-6-trifluoro- methyl-4-pyridine- methanol	GC	3% OV-17 on silanized Chromo- sorb W, 100—200 mesh	Nitrogen, 30 ml/min	FID or ⁶³ Ni ECD	1 ng/ml (ECD); 100 ng/ml (FID)	Blood	Derivatization with hexamethyldisilane and trimethylchlorosilane; column programmed 160—250°C (1°C/min), injector and detector 250°C	302
	HPLC	μBondapak C ₁₈	0.005 M <i>Low</i> UV PIC B 8 in methanol—water (70:30), 1 ml/min	UV 222 nm	Below 10 ng/ml	Plasma, blood		303
	TLC	Silica gel	Heptane— <i>n</i> -butanol—glacial acetic acid (8:1:1) (six serial developments); <i>n</i> -butanol— glacial acetic acid—water (66:17:17)	UV 254 nm, ¹⁴ C autoradi- ography		Urine, faeces, blood, expired air		304
	TLC	Silica gel	Dichloromethane—methanol— acetic acid (98:8:2); dichloro- methane—methanol—conc. ammonium hydroxide (80:19:1); chloroform—methyl ethyl ketone—methanol (20:65:16); benzene—methyl acetate—methanol (40:40:20)	¹⁴ C Liquid scintillation counting		Blood, urine, faeces		305

(Continued on p. 444)

TABLE 10 (continued)

Drugs separated	Method	Stationary phase	Mobile phases or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Mefloquin + metabolite	TLC	Silica gel	Dichloromethane-methanol-acetic acid (80:10:10)	UV 300 nm, densitometry	100 ng/ml	Blood, plasma		306
Maloprim, see Pyrimethamine	GC	3% SP-2401 on Supelcoport, 80-100 mesh	Nitrogen, 46 ml/min	^{63}Ni ECD	1-2 ng/ml	Blood	Derivatization with heptafluorobutyric anhydride; column 235°C, injector 200°C, detector 300°C	307
Primaquine	GC	3% OV-17 on Anachrom ABS, 11.0-12.0 mesh Partisil ODS III RP (10 μm)	Carrier gas, 30 ml/min	Nitrogen-sensitive detection, FID			Column 265°C	308
	HPLC		Water-acetonitrile-methanol (60:30:10) cont. 5 \cdot 10 $^{-4}$ M octanesulfonic acid, 1.5 ml/min	UV	1 ng/ml	Plasma, urine	Mobile phase buffered to pH 3.5	309
Primaquine + metabolite [8-(3-carboxy-1-methylpropylamino)-6-methoxyquinoxine]	HPLC	μ Bondapak C ₁₈ RP (10 μm); pre-column Whatman C ₁₈	6.6 g K ₂ HPO ₄ , 8.4 g KH ₂ PO ₄ , 2.4 l methanol, 1.6 l water, cont. 4 g N,N-dimethyloctylamine, 1.0 ml/min	UV 254 and 280 nm; liquid scintillation counting	0.05 $\mu\text{g}/\text{ml}$	Plasma, lungs, adrenal glands, liver, faeces		308, 310
	HPLC	μ Bondapak CN RP (10 μm)	Acetonitrile-0.08 M citrate buffer pH 5.0 (77:23), 1 ml/min	ECD	0.2 ng	Plasma, urine		311
Primaquine + metabolites	TLC					Urine		312
Pyrimethamine, metoprine	GC	10% OV-17 on Chromosorb W HP, 100-120 mesh	Nitrogen, 35 ml/min	ECD	Below 5 ng/ml	Plasma	Oven 235°C, injector 300°C, detector 350°C; when internal standard is eluted, the temperature was programmed to 280°C (16°C/min)	313
	GC	3% OV-17 on Chromosorb 750 80-100 mesh (5 μm)	Argon-methane (9:1), 60 ml/min (plasma), 50 ml/min (urine)	^{63}Ni ECD		Plasma, urine	Column 235°C, detector 350°C (plasma), column 220°C (urine)	314
Pyrimethamine, dapone (maloprim), monadecyldapsone, metoprine	HPLC	Spherisorb S5W (5 μm)	Diisopropyl ether-methanol-21% ammonia (96:4:0.1), 2 ml/min	UV 254 nm	5 ng per injection	Plasma		315
	LC	μ Bondapak C ₁₈ (10 μm)	MeOH-acetonitrile-water (25:15:60) cont. 0.005 M pentaneulphonic acid, 1.5 ml/min	UV 254 nm	5 ng/ml	Plasma		316

HPLC	LiChrosorb Si 60 (5 μ m)	Methanol—acetonitrile—25% ammonia—diisopropyl ether (6:25:0.1:71)	Fluorescence at 290/345 nm	10 ng/ml	Plasma	317
HPLC	μ Bondapak C ₁₈ (10 μ m)	Methanol—acetonitrile—water (25:1:60) cont. 0.005 M 1-pentanesulphonic acid, 1.5 ml/min.	UV 254	5 ng/ml pyrimethamine; 50 ng/ml sulphadoxine; 5 ng/ml N-acetyl sulphadoxine	Plasma	318
Pyrimethamine, see also Chloroquine						
Quinine, quinidine, metabolites (see also Chloroquine)	GC, GC-MS	Silanized glass coil column, 1% SE-30 on Gae-Chrom Q, 100–120 mesh	MS		Urine	319
HPLC	μ Bondapak C ₁₈ , RP	(A) Water—acetic acid (99:1); (B) water—acetonitrile—acetic acid (40:5:9:1)	UV 254 nm	Proportion of B varied from 10 to 85%	Urine	
HPLC	μ Bondapak C ₁₈ , RP	(A) Water—acetic acid (99:1); (B) water—acetonitrile—acetic acid—tetrahydrofuran (40:59:1:0.1), 1.8 ml/min	UV 254 nm	Proportion of B varied from 10 to 80% (semipreparative method)	Urine	
Sulphadoxine, see Pyrimethamine						
DL-threo-(2-Piperidyl)-2-trifluoromethyl-4-(4-trifluoromethylphenyl)-4-pyridinomethanol	TLC	Silica gel	<i>n</i> -Butanol—acetic acid—water (65:7:17); benzene—methanol (3:1)	¹⁴ C Autoradiography, UV 254 nm	Plasma, red blood cells, urine, faeces, expired air	320

TABLE 11
ANTIPARASITICS, ANTIPROTOZOAL DRUGS (EXCLUDING ANTIMALARIALS)

Drugs separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Avermectins	HPLC	Zorbax ODS	Acetonitrile-methanol-water (65:35:7), 0.8 ml/min	UV 254 nm	2 ng/ml	Plasma		321
Azanidazole (Tridazole)	HPLC	Partiai 10 ODS/2 RP (10 μ m)	52% Methanol in eq. KH ₂ PO ₄ , (0.1%), 2 ml/min	UV 368 nm	10 ng/ml	Plasma, urine		322
3a,4,5,6,7,7a-Hexahydro-3-(1-methyl-5-nitro-1H-imidazol-2-yl)-1,2-benzisoxazole	GC-MS	SE-30 capillary column	Helium, 1 ml/min			Liver	Derivatization with N,O-bis(trimethylsilyl)trifluoroacetamide; column 220°C	323
	TLC	Silica gel	Toluene-ethyl acetate (3:1); dichloromethane-methanol (9:1); trimethylpentane-2-propanol (4:1)	¹⁴ C Liquid scintillation counting		Liver		323
<i>Iodochlorhydrxyquin, see Antibacterials</i>								
5-Isopropyl-1-methyl-2-nitro-1H-imidazole (DL 347)	GC	3% OV-25 on Chromosorb G HP, 80-100 mesh	Helium, 40 ml/min	FID or ¹⁴ C counter detection		Urine	Column programmed 100°C to 230°C (3°C/min), inlet and detector 250°C	324
	GC-MS	3% OV-25 on Chromosorb G HP, 80-100 mesh	Helium, 15 ml/min	MS		Urine	Column programmed 120°C to 250°C (4°C/min), inlet 280°C, manifold 240°C, gas inlet 250°C, ion source 200°C	324
	TLC	Silica gel	Benzene-methanol (95:5 or 80:20); chloroform-methanol (99:1, 95:5 or 80:20)	¹⁴ C Autoradiography		Urine		324
Metronidazole (see also Ornidazole)	GC	3% OV-11 on Supelcoper, 100-120 mesh	Nitrogen, 50 ml/min	⁶³ Ni ECD	50 ng/ml	Blood	Derivatization with (trimethylsilyl)trifluoroacetamide; column 180°C, injector 150°C, detector 250°C	325
Metronidazole, misonidazole	HPLC	μ Bondapak C ₁₈ RP	8% Acetonitrile in 10 ⁻³ M phosphate buffer pH 4.0, 2 ml/min	UV 324 nm	0.5 μ g/ml	Plasma, urine		326

Metronidazole, tinidazole	HPLC (5 μm)	Spherisorb S5 ODS (5 μm)	0.01 M Phosphate buffer pH 5.5 UV 320 nm mixed with 15% acetonitrile, 1 ml/min	25 ng/ml metronidazole faeces	Plasma, faeces	327
Metronidazole, tinidazole	HPLC μBondapak C ₁₈	μBondapak C ₁₈	7% Acetonitrile in 20 mM acetate buffer pH 4.0; 1.5 ml/min for metronidazole; 2.0 ml/min for tinidazole	0.5 μg/ml metronidazole; 2.0 μg/ml tinidazole	Plasma	328
Metronidazole + metabolites (hydroxymetronida- zole-1-acetic acid)	HPLC (10 μm)	μBondapak C ₁₈ , RP (10 μm)	0.005 M KH ₂ PO ₄ pH 4.5—meth- UV 234 nm anol—tetrahydrofuran (82.6:16.5:0.9, 1.4 ml/min)	0.05 μg/ml	Plasma, urine	329
Metronidazole + metabolites	HPLC μBondapak C ₁₈ , RP	μBondapak C ₁₈ , RP M KH ₂ PO ₄ , pH 4.0 (4:3:93)	Methanol—acetonitrile—0.005 M KH ₂ PO ₄ , pH 4.0 (4:3:93)	6.8 μg/ml	Plasma	330
Metronidazole + major metabolites [1-(2-hydroxyethyl)-2- hydroxymethyl-5- nitroimidazole (I); 2-methyl-5-nitroim- dazole-1-acetic acid (III)]	HPLC HPLC HPLC	Spherisorb 5-10 ODS	0.1 M (NH ₄) ₂ HPO ₄ —methanol (5:1), 0.5 ml/min	UV 328 nm 20–50 ng/ml (plasma), 1 μg/ml (urine)	Plasma, urine	331
Metronidazole HPLC HPLC	LiChrosorb RP 8	LiChrosorb RP 8	Ethanol—water (1:9), 2 ml/min	UV 371 nm 1 μg/ml	Plasma Biological fluids	332 333
Metronidazole, misonidazole + metabolites	HPTLC RP-18 (Merck)	n-Hexane—acetone—ethanol (19:6:1)	UV 320 nm; scanning denatometry	0.5 μg/ml	Serum, urine	335
Misonidazole + desmethyl metabolite	HPLC (10 μm)	μBondapak C ₁₈ , RP (10 μm)	19% Methanol—water, 2 ml/min	UV 313 nm Below 2 μg/ml	Plasma, urine, tissue homogenates	336
Misonidazole + desmethyl metabolite (see also Metronidazole)	HPLC RP-18 (10 μm)	LiChrosorb 10 RP-18 (10 μm)	Methanol—water (20:80), 2 ml/min	UV 313 nm 1.4 μg/ml misonidazole; 0.7 μg/ml des- methyl derivative	Plasma	337

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TABLE 11 (continued)

Drugs separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Misonidazole + desmethyl metabolite	HPLC	Partisil PXS 5/25 ODS	19% Methanol—water, 2 ml/min			Rat liver	Details see ref. 320	338
Misonidazole + desmethyl metabolite	HPLC	Ultrasphere ODS (5 μ m), Alter, Varian MCH:10 (1.0 μ m); Varian MCH:5 (5 μ m); Varian MCH:N-cap 5 (5 μ m)	1 mM Potassium phosphate buffer pH 4.0—acetonitrile (93:7), 1.5 ml/min	UV 323 or 313 nm	0.4 μ g/ml misonidazole; 0.2 μ g/ml desmethyl-misonidazole	Plasma, serum		339
Misonidazole + desmethyl metabolite (1-methyl-2-nitro-5-vinyl imidazole), cyclophosphamide, 5-fluorouracil	TLC	Silica gel	Ethyl acetate—dichloromethane—methanol (5:3:1 or 15:10:1); dichloromethane—methanol (7:5:1); chloroform—ethyl acetate—ethanol (7:5:7:5:1)	Densitometry at 320 nm	1 μ g/ml	Plasma	Sep-Pak C ₁₈ extraction	340
Nitrimidazole	HPLC	μ Bondapak C ₁₈ (10 μ m)	Methanol—water (30:70) cont. PIC reagent B-7, 2 ml/min	UV 313 nm	5 μ g/ml	Blood		341
Ornidazole, metronidazole	GC	3% OV-11 on Supelcoport, 100–120 mesh	Nitrogen, 50 ml/min	⁶³ Ni ECD	50 ng/ml	Blood	Derivatization with N,O-bis(trimethylsilyl)trifluoroacetamide; column 180°C, injector 150°C, detector 250°C	325
Ornidazole + metabolites: 1-(chloromethyl)-2-hydroxymethyl-5-nitroimidazole-1-ethanol; 3-(2-methyl-5-nitroimidazole-1-yl)-1,2-propanediol	HPLC	μ Bondapak C ₁₈ , RP (10 μ m)	Water—ethanol (1:9), 2 ml/min	UV 318 nm	0.2 μ g/ml	Plasma	Also possible LiChrosorb RP-2, RP-8 or RP-2, 342	342
Salicyhydroxamic acid	HPLC	Magnusphere C ₁₂ RP (5 μ m)	0.043 M NH ₄ H ₂ PO ₄ —methanol (70:30) pH 2.0, 1.2 ml/min	UV 300 nm	0.1 μ g/ml	Plasma		343
Satravidazole	GC	3% OV-11 on Supelcoport, 100–120 mesh	Nitrogen, 40 ml/min	⁶³ Ni ECD	50 ng/ml	Blood	Derivatization with N,O-bis(trimethylsilyl)trifluoroacetamide; column 280°C, injector 220°C, detector 300°C	325

Secnidazole	GC	3% OV-11 on Supelcort, 100–120 mesh	Nitrogen, 50 ml/min	^{63}Ni ECD	50 ng/ml	Blood	Derivatization with N,O-bis(trimethylsilyl)trifluoroacetamide; column 180°C, injector 150°C, detector 250°C	325
Thnidazole (see also Metronidazole)	GC	3% OV-11 on Supelcort, 100–120 mesh	Nitrogen, 45 ml/min	^{63}Ni ECD	50 ng/ml	Blood	Derivatization with N,O-bis(trimethylsilyl)trifluoroacetamide; column 280°C, injector 200°C, detector 300°C	325
	GC	3% OV-11 on Gas-Chrom Q, 80–100 mesh	Nitrogen, 20 ml/min	NPD	20 ng/ml	Plasma, tissue	Column 215°C, injector and detector 245°C	344
Trnidazole + two metabolites: ethyl[2-(2-hydroxy- methyl-5-nitro-1- imidazolyl)ethyl]- sulphone, its O-glu- curonide conjugate	HPLC RP	μ Bondapak/Phenyl pH 7.0—methanol (86:14), 2 ml/min	0.05 M KH ₂ PO ₄ —NaOH buffer Hexane—chloroform—ethanol (90:15:0.5), 1 ml/min Chloroform—acetic acid— methanol (18:2:1:0:0.8)	UV 313 nm UV 315 nm Fluorescence at 366 nm	0.1 $\mu\text{g}/\text{ml}$ 0.2 $\mu\text{g}/\text{ml}$	Plasma		345
	HPLC	ETH Permaphase (25–37 μm) Silica gel				Plasma		346
	HPLC					Plasma		102

TABLE 12
ANTIPARKINSONICS

Drugs separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Amantadine (Symmetrel)	GC	5% Apiezon L on Gas-Chrom Q, 100-120 mesh	Nitrogen, 60 ml/min	FID	Below 0.1 $\mu\text{g}/\text{ml}$ (plasma); 4 $\mu\text{g}/\text{ml}$ (urine)	Plasma, urine	Silanized glass; column and oven 150°C, injector 250°C, detector 300°C	347
	GC-MS	3% OV-225 on Gas-Chrom Q; 3% OV-17 on Chromosorb W HP;	Carrier gas, 40 ml/min	MS		Plasma	Derivatization: N-acetyl or isothiocyanate derivatives, columns 170°C, 180°C or 160°C	348
	GC-MS	3% OV-101 on Gas-Chrom Q	Helium, 30 ml/min	MS			Column 190°C, ion source 280°C	
	GC	3% OV-225 on Gas-Chrom Q 5% SE-30 on Chromosorb W HP, 80-100 mesh	Argon-methane (90:10), 75 ml/min	ECD	Below 10 ng/ml	Plasma, urine	Silanized glass; derivatization with trichloroacetyl chloride; column 200°C, injector 250°C, detector 300°C	349
	GC	10% Apiezon L and 2% KOH on Chromosorb W AW, 80-100 mesh	Nitrogen, 50 ml/min	FID	0.5 $\mu\text{g}/\text{ml}$	Urine	Column 145°C, detector 170°C, injector 180°C	350
Benserazide	TLC + Elpho	Cellulose; first direction Elpho, second direction TLC	ad (1) Aq. buffer pH 4.4 (1/6 M in pyridine, 1/3 M in acetic acid), 18 mA, 80 min; ad (2) 2-propanol-water— acetic acid (50:50:5)	^{14}C Autoradiog- raphy		Plasma, urine		351
Benztropine	GC-MS	1% OV-17 on Gas-Chrom Q, 100-200 mesh	Helium, 20 ml/min (conditioned)	Electron-impact ionization	5 ng/ml	Urine, plasma	Silanized glass column; column 230°C, flask heater 240°C, ion source 270°C	352
Benztropine, trihexyphenidyl	LC	Micropak CN (10 μm)	Acetonitrile or methanol-aq. 0.005-0.2 M ammonium acetate (90:10), 2.0-2.5 ml/min	UV and electro- chemical detection	10 ng/ml	Blood, plasma		209
Biperiden	GC	OV-101 capillary column	Helium, 2 ml/min	Nitrogen-sensitive detection	25 pg/ml	Serum	Column 215°C, injector 250°C, detector 300°C	353

Bromocriptine (Parlodol)	GC	1% OV-17 on Celite JI CQ, 100–120 mesh	Argon–methane (9:1), 50 ml/min	Detection in a pulsed mode (150 μ sec) MS	0.5 ng/ml	Plasma	Derivatization with hexamethyl- dilazane; column 245°C, injec- tor 250°C, detector 350°C
	GC-MS	3% OV-101 on Celite JI CQ, 100–120 mesh	Helium, 20 ml/min		1.0 ng/ml	Plasma	Column 180°C, injector 200°C, molecule separator 260°C, ion source 290°C
HPLC		μ -Bondapak C ₁₈	Methanol–water (65:35), cont.	UV 254 nm	10 ng/ml	Plasma	
			0.01 M 1-heptanesulphonic acid, 1.5 ml/min				
Carbidopa, see Levodopa							
Deprenyl (Jumex)	GC	Carbowax 20 M glass capillary column	Nitrogen, 2.5 ml/min	NPD	400 pg/ml	Plasma	Column programmed from 70°C to 105°C (20°C/min), then to 170°C (4°C/min) injector 200°C, detector 280°C
Deprenyl + metabolites	TLC	Silica gel	(1) Chloroform–methanol– borate buffer pH 6.5 (7:6:1); (2) butanol–acetic acid–water (3:1:1); (3) <i>tert</i> -butanol– ammonia–water–methanol (20:1:4:2); (4) phenol–water (8:2)	UV 254 nm		Urine	
Levodopa (L-dopa) + metabolites, (dopamine, norepinephrine)	GC-MS	3% OV-1 on Supelcortor, 80–100 mesh	Nitrogen, 30 ml/min	⁶³ Ni ECD, MS	0.5 pg (metabolites)	Brain extracts	Silanized glass columns; catecholamines converted to N-2,6-dinitro-4-trifluoromethyl- phenyl-O-trimethylsilyl derivatives
Levodopa, carbidopa, 3,4-dihydroxyphenyl- acetic acid	HPLC	Spherisorb ODS (5 μ m)	100 mM NaH ₂ PO ₄ , 20 mM citric acid, 1.25 mM sodium 1-octanesulphonic acid, 0.15 mM sodium EDTA in 8% methanol, 1 ml/min	Electrochemical detection	15 ng/ml	Blood, plasma	
	HPLC	Ultrasphere-octyl Altex (5 μ m)	Citrate–phosphate buffer pH 3.1 cont. 6.5 mM 1-octane- sulphonic acid and 1.4% methanol; disodium salt of EDTA was added to final conc. 2 mM, 1.2 ml/min	Amperometric detection	25 ng/ml	Plasma	

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TABLE 12 (continued)

Drug separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Orphenadrin (Disipal) + metabolite	GC	3% OV-17 on Chromosorb W AW DMCS HP, 80—100 mesh	Nitrogen, 40 ml/min	FID	0.8 µg/ml	Serum	Silanized glass column; column 250°C, injector 260°C, detector 270°C	360
Orphenadrin, diphenhydramin	GC	Capillary fused- column DB-1	Helium, 1.2 bars		2 ng/ml	Serum	Oven 50°C (1 min), then to 180°C (5°C/min) and to 210°C (6°C/min), injector 205°C	361
Procyclidine (Kemaditen)	GC	3% OV-17 on Chromosorb W HP, 60—80 mesh	Nitrogen, 20 ml/min	FID		Plasma	Methyl derivatives; oven 140°C 2 min, then programmed to 230°C, 4°C/min, injector 230°C, detector 280°C	362
	GC	5% OV-17 on Chromosorb W HP, 100—120 mesh	Helium, 50 ml/min	NPD	20 ng/ml	Plasma, urine	Manifold 32°C, oven 246°C, injector 310°C	363
Procyclidine + metabolite [1-(3-hydroxycyclo- hexyl)-1-phenyl-3- (1-pyrrolidinyl)-1- propanol)]	GC-MS	2% OV-101 and 0.2% Carbowax 20 M on Chromo- sorb W HP, 100—120 mesh	Helium, 20 ml/min	MS		Urine	Column programmed 130°C to 230°C (8°C/min), injector 150°C	364
	TLC	Silica gel					Chloroform—methanol—conc. ammonium hydroxide (50:50:1); then transfer to second plate, methanol—conc. ammonium hydroxide (100:1:5)	Preparative TLC

TABLE 13
ANTITUSSIVES

Drugs separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Bazitramide (Burgodin) + two metabolites [mainly 1-(4-piperidinyl)-1,3- dihydro-2H-benzimi- dazol-2-one]	HPLC	LiChrosorb RP-2 (10 μ m)	Acetonitrile-methanol-isopropylamine (930:70:5); 6.9 ml/min/sec	UV 250 nm	Below 1 μ g/ml	Urine	Basic metabolite of droperidol can be separated	365
Bromhexine (Adamoxine)	GC	4% SE-30 on Gas-Chrom Q, 100-120 mesh	Argon-methane (95:5), 28 ml/min	15 mCi α -Ni ECD	About 1.0 ng/ml 1.0 ng/ml	Plasma	Silanized glass column; derivatization with trifluoroacetic anhydride; oven 255°C, injector 280°C, detector 330°C	366
	HPLC	μ Bondapak C ₁₈ (10 μ m); pre-column Nucleosil C ₁₈ (10 μ m) Capillary column OV-17	Acetonitrile-methanol-0.01 M phosphate buffer pH 7.0 (40:40:20 for plasma and 41:41:18 for urine) Helium, 2 ml/min	UV 254 nm NPD	5 ng/ml (plasma); 2.5 ng/ml (urine) 0.5 ng/ml	Plasma, urine	Column programmed from 60°C to 140°C (10°C/min), then to 230°C (5°C/min), injector 240°C, detector 260°C	367
GC-MS	Capillary column OV-17	Helium, 2 ml/min	SIM	Diazotation and coupling, or fluorogenic labeling with fluorescein amine and fluorimetry at 510 nm	250 ng/ml	Plasma	Column programmed from 60°C to 230°C (10°C/min), injector 260°C	368
TLC	Silica gel	Ethyl acetate-acetic acid-water (60:15:15)	Nitrogen-sensitive detection	5 ng/ml	Plasma			369
Codeine	GC	1% OV-17 on Chromosorb W HP, 100-120 mesh	Helium, 30 ml/min			Column 235°C, injector 260°C, detector 300°C		370
Codeine, other opiates + metabolites	GC-MS	3% OV-210, OV 101, OV-17, 5CP on Gas-Chrom Q, 100-120 mesh	Nitrogen, 20 ml/min	FID, MS	0.01 μ g/ml	Urine	Trimethylsilyl derivatives; column programmed from 170°C to 260°C (10°C/min) (36-cm column) or 220°C to 260°C (10°C/min) (183-cm column), injector 190°C, detector 275°C	371

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TABLE 13 (continued)

Drugs separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Codine, other opiates	GC	Glass-capillary column (borosili- cate, deactivated with N-cyclohexyl- 3-acetidenoil); or Carbowax 20 M, OV-1, OV-17, OV-225 Carbowax 20 M fused silica	Helium, 35 ml/min	ECD	5 ng/ml	Plasma, blood	Derivatization with pentafluoro- propionic anhydride; column 220°C, injector 250°C, detector 300°C	372
Codine, other opiates	GC, GC-MS	3% Silia-5CP (A), 3% Silia-10C, 3% OV-225 (B) or 3% OV-17 (C) on Gas-Chrom Q, 100-120 mesh μPorasil (10 μm); pre-column Vydac 101 SC C ₁₈ , μBondapak; pre-column Co-Pell ODS Silica gel	Nitrogen, 60 ml/min	FTD, MS	10 μg/ml	Urine	Trimethylsilyl derivatives; column: codeine underivatized 250°C (A), 240°C (B, C), silylated 250°C (A), 210°C (B), 240°C (C); injector 275°C detector 275°C	373
HPLC		Dichloromethane—methanol— 33% ammonia (90:10:0.1), 1.5 ml/min	UV 254 nm	50 ng/ml	Plasma			374
HPLC		Methanol—water (21:79) cont. 1.5 g phosphoric acid, 2 ml/min	Fluorescence at 213/210 nm	4 ng/ml	Plasma			375
HPTLC		Chloroform—acetic acid— methanol (18:2:1:0.8)	Fluorescence at 366 nm		Plasma			102
2',4'-Dimethyl-6'- methoxy-5-(2-methyl- piperidyl)-propion- aldehyde (or K-242) + metabolites	HPLC	Spherisorb CN (5 μm)	30% Acetonitrile in 1 mM sodium phosphate pH 4.0, 1.5 ml/min	UV 214 nm	2 ng/ml	Plasma		376
Eprazinone (Etagapan, Mucitux) + metabolites	GC-MS	3% OV-17 on Gas-Chrom Q, 100-120 mesh	Helium, 20 ml/min	MS		Urine	Identification of metabolites; open programmed 100°C to 280°C (1.5°C/min), ion source 250°C, separator 260°C	377
TLC		Silica gel	Methanol—ammonium hydroxide (100:1); cyclo- hexane—diethylamine (9:1); chloroform—acetone (85:15); benzene—acetyl acetate—di- ethylamine (7:2:1); benzene— ethanol—ammonium hydroxide (80:20:1)	Chemical (Dragendorff)	0.5 μg			

Guiphenesin + metabolite [α -(2-methoxyphenoxyl)-lactic acid]	HPLC	LiChrosorb RP 8 (5 μ m), 30°C	Methanol—0.01 M citrate buffer UV 275 nm pH 6.5 (40:60), 1 ml/min	Below 5 μ g/ml	Plasma	378
Hydrocodone	GC	3% OV-7 on Supelcoport, 100—120 mesh Capillary column with SE-30	Argon—methane (95:5), 40 ml/min	^{63}Ni ECD 1 ng/ml	Serum	379
	GC		Helium	Nitrogen-sensitive detector	0.2 ng/ml	380
Noscapine, noscapine acid	LC	LiChrosorb Si 60 (5 μ m) Spherisorb S 5 ODS	Hexane—methanol—chloroform—diethylamine (86.5:10:1:3.4:0.034), 1 ml/min 0.005 M Pentanesulfophonic acid in methanol—water—acetic acid—triethylamine (40:53:6:1), 1 ml/min	UV 310 nm 5 ng/ml (straight phase); 15 ng/ml (RP)	Plasma	381
Racemethorphan (dextromethorphan)	GC	3% OV-25 on Supelcoport, 80—100 mesh	Argon—methane (95:5), 50—55 ml/min	^{63}Ni ECD 1 ng/ml	Serum	382
	GC	2% OV-101 on Chromosorb W HP, 100—120 mesh		Nitrogen-sensitive detector	Below 1 ng/ml	383
Racemethorphan + metabolites (dextrophorphan, 3-hydroxy- δ - ϵ , 13 α , 14 α -morphinan, 3-methoxy- δ - ϵ , 13 α , 14 α -morphinan)	HPLC	Spherisorb phenyl RP (5 μ m)	10 mM Monobasic potassium phosphate—acetoneitrile (45:55) pH 4.0, 1.2 ml/min	UV 280 nm 0.017—0.09 μ g/ml (base); 0.11—0.21 μ g/ml in hydrolyzed or non-hydrolyzed urine	Urine	384
Zipeprol + metabolites	GC	(1) Apiezon L on Chromosorb W AW DMCS, 80—100 mesh; (2) 3% OV-17 on Chromosorb S AW DMCS, 80—100 mesh; (3) 1% OV-1 on Supelcoport, 100—120 mesh; (4) 2% OV-1 + 2% KOH on Chromosorb W 100—120 mesh; (5) 3% SE-30 on Chromosorb W, 100—120 mesh; (6) 3% SP-2250 on Supelcoport, 100—120 mesh	Nitrogen FID	Urine	Derivatization; trifluoroacetyl- and silyl derivatives; TLC- preparative isolation of individual metabolites	385

TABLE 14
ANTIULCER DRUGS
Check also Antihistamines for completeness.

Drugs separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Aldioxa	HPLC	Zorbax ODS (5–6 μ m)	Acetonitrile—water (27:73); after 3 min of the appearance of the peak the mobile phase was changed to methanol—water (70:30) (18 min), then again changed to acq. acetonitrile, 1.0 ml/min	UV 241 nm	100 ng/ml	Plasma	Hydrolysis of aldioxa to allantoin and its conversion to xanthylallantoin, which is determined (derivationization with xanthylidrol). Sep-Pak C ₁₈ cartridges for extraction	386
Cimetidine	HPLC	Partisil 10 ODS (10 μ m)	Acetonitrile—water—ammonium hydroxide (1.000:50:1), 2.5 ml/min	UV 228 nm	0.1 μ g/ml, using 750 μ l of plasma	Plasma		387
	HPLC	Ultrasphere ODS (5 μ m); pre-column Partisil RP-2, (30–40 μ m) before pump and injector; LiChrosorb RP-2 (10 μ m) before analytical column	10 mM Phosphate buffer (pH 8.0)—methanol (80:20), 1.7 ml/min	UV 220 nm	0.1 μ g/ml	Plasma, urine		388
	HPLC	Partisil 10 ODS-3	Acetonitrile—10 mM potassium phosphate pH 4.8 (7:39), 2.0 ml/min	UV 228 nm	0.1 μ l/ml (200 μ l of biological fluid was used)	Serum, urine	Interference of procainamide and tolazamide	389
	HPLC	MicroPak CN-10 (10 μ m)	Acetonitrile (15 min), 1 ml/min, UV 238 nm then acetonitrile—0.01 M NaH ₂ PO ₄ (60:50)	UV 238 nm	0.1 μ g/ml (0.05 μ g/ml)	Plasma		390
	HPLC	Partisil 10 ODS	1/15 M Monoethic potassium phosphate—1/15 M dibasic sodium phosphate—methanol, (8:15:1.00:1.85), 2.5 ml/min	UV 228 nm	0.1 μ g/ml	Urine, plasma	Sep-Pak C ₁₈ cartridges for extraction	391
	HPLC	LiChrosorb Si-100 (10 μ m)	Acetonitrile—methanol—water—25% ammonium hydroxide (250:30:10:0.4), 1 ml/min	UV 228 nm	0.03 μ g/ml	Plasma		392
	HPLC	LiChrosorb RP-18 (5 μ m)	Methanol—0.01 M ammonium carbamate pH 8.9 (40:60), 1.2 ml/min	UV 220 nm	0.08 μ mol/l	Plasma		393

Cimetidine + metabolite (cimetidine sulph- oxide)	HPLC	LiChrosorb Si 60 (5 μ m)	Acetonitrile—methanol—water— ammonium hydroxide (sp. gr. 0.88) (250:20:6:1.5), 1 ml/min	UV 228 nm 0.05 μ g/ml cimetidine; 0.2 μ g/ml cimetidine sulphoxide 0.05 μ g/ml	Plasma, urine	394
	HPLC	Zorbax Sil; pre-column Whitman HC Pellicol RP Radial-Pak A Waters Assoc.	Acetonitrile—methanol—water— ammonium hydroxide (1000:50:50:2) pH 10.5, 3 ml/min 1% Triethylamine and 5% aceto- nitrile in water (pH 3.0, phosphoric acid), 3 ml/min	UV 228 nm 25 ng/ml	Serum, plasma, urine	395
	HPLC	μ Bondapak C ₁₈	Methanol—5 mM KH ₂ PO ₄ , pH 2.8 (10:90), 20 ml/min Acetonitrile—methanol— water—ammonia (1.000:200:20:5)	UV 228 nm Below 0.1 μ g/ml	Serum, plasma	396,
	HPLC	LiChrosorb Si 60 or Partisil 5 (5 μ m)	Acetonitrile—conc. ammonia— water (1000:2.5:20), 1.6 ml/min Methanol—0.075 M mono- basic ammonium phosphate pH 2.5 (15:85)	UV 228 nm 0.05 μ g/ml	Serum Blood	397
	HPLC	RP Ultrasphere- octyl	Ethanol—acetic acid—water Photometry at 636 nm (Folin— Ciocalteu reagent)	UV 220 nm 0.02 μ g/ml	Plasma, gastric fluid	400
	TLC	Silica gel	Ethy acetate—methanol— ammonium hydroxide (sp. gr. 0.88) (8:1:1); acetone— ethyl acetate—ammonium hydroxide (sp. gr. 0.88) (6:2:2)	Photometry at 636 nm (Folin— Ciocalteu reagent) UV, ¹⁴ C auto- radiography	Blood, urine	401
	TLC	Silica gel	Ethy acetate—methanol— ammonium hydroxide (sp. gr. 0.88) (8:1:1 or 8:1:1); dichloromethane—methano saturated with ammonium hydroxide (4:1)	Urine, faeces	398	
Cimetidine, metiamide + metabolites	TLC	Silica gel	¹⁴ C, ³ H Auto- radiography	Plasma, urine	405	

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TABLE 14 (continued)

Drugs separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Cimetidine	HPTLC	Silica gel	Chloroform—acetic acid—methanol (18:2:1:0.8)	Fluorescence at 366 nm		Plasma		102
N-(2-(Diisopropylaminoethyl)-N-(4,6-dimethyl-2-pyridyl)-N,N'-dimethylurea	GC	1.5% or 3% OV-17 on Gas-Chrom Q, 100–120 mesh	Hydrogen, helium, air, 3.5, 20, 100 ml/min or 60, 46, 550 ml/min	Nitrogen-sensitive detection, FID		Plasma, urine	Derivatization with bis(trimethylsilyl)trifluoroacetamide, trifluoroacetic anhydride; column 210, 226 or 245°C, injector 260°C, interface 275°C, oven 180–200°C, source 270°C, separator 270°C, injector 230°C	406
	GC-MS	1% OV-17 on Supelcort, 80–100 mesh Silica gel	Helium, 30 ml/min	MS				
	TLC		Chloroform saturated with ammonium hydroxide—methanol (1:9); acetone—conc. ammonium hydroxide; chloroform—methanol—acetic acid (4:7.5:47.5:5); benzene—dioxane—conc. ammonium hydroxide (60:35:5); upper phase: benzene—dioxane—conc. ammonium hydroxide (10:80:10); methanol—conc. ammonium hydroxide (100:1:5)	¹³ C Radio-scanning and UV				
Geranylgeranyl-acetone	GC-MS	3% OV-17 on Gas-Chrom Q, 100–120 mesh	Helium, 30 ml/min	MS	1 ng/ml	Serum	Derivatization with O-(2,3,4,5,6-5-fluorobenzyl)hydroxyamine; column 270°C, injector and separator 300°C, ion source 330°C	407
Ormetidine, see Antihistamines								
Pirenzepine (Gastropeptine), ranitidine	HPLC	Lichrosorb RP-8 (5 µm) (for Gastropeptine); 23°C; Spherisorb ODS (5 µm) (for ranitidine) 5:1°C	Methanol—0.05 M phosphate buffer pH 7.0	UV-VIS spectrophotometry, ranitidine 320 nm, gastropeptine 283 nm		Blood		408
Ranitidine + metabolites	HPLC	Spherisorb QDS (5 µm), 45°C	Methanol—0.0005 M aq. NaH ₂ PO ₄ + 0.005 M aq. sodium dodecyl sulphate (60:40), 1 ml/min	UV 320 nm	0.8 µg/ml	Urine, serum		409
	HPLC	Spherisorb S5 CN	Methanol—2-propanol—5 M ammonium acetate (50:50:1), 0.5 ml/min	UV 320 nm	1 µg/ml	Urine	Combination with MS-SIM technique	410
Ranitidine (see also Pirenzepine)	HPLC	RP			10 ng/ml	Plasma, Urine, faeces, other biological material		411 412

TABLE 15
ANTIVIRAL COMPOUNDS

Drugs separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Acyclovir	HPLC	Zorbax ODS silica (5 μ m)	0.005 M Sodium acetate and 0.0025 M heptanesulphonic acid, sodium salt (pH 6.5)	UV 254 nm	10 μ M	Plasma, urine		413
Amantadine, see Antiparkinsonics								
Arlidone	GC	3% OV-1 on Gas-Chrom Q, 100-120 mesh	Argon-methane (93:7), 60 ml/min	ECD	1.4 ng/ml (urine); 6.4 ng/ml (plasma); 12.6 ng/g (faeces)	Plasma, urine, faeces	Derivatization with O-(2,3,4,6-penta(hydrobenzyl)hydriodine; column 275°C (285°C for faecal analysis), injector, detector 300°C	414
Arlidone + metabolites	HPLC	Lichrosorb RP-8 or Lichrosorb RP-18	Acetonitrile-water-98% formic acid (1.000:20:2 or 10:90:0.5; water-formic acid (100:0.1), 2 ml/min	UV 280 nm or radioactivity measuring	Below 0.12 μ g/ml	Blood	Gradient for metabolite study: water-acetonitrile-formic acid (100:0.1 to 0:100:0.1)	415
	TLC	Silica gel	<i>n</i> -Hexane-diethyl ether (1:1); chloroform-methanol (99:1); chloroform-ethanol (80:20)	¹⁴ C Radioactivity scanning				
Cytarabine (cytosine arabinoside)	GC	3% SE-30, 3% OV-17, 3% OV-210, all on Gas-Chrom Q, 100-120 mesh	Nitrogen, 30 ml/min	NPD	500 pg	Plasma	Derivatization: acetyl methyl derivatives, trimethylsilyl di-methyl derivatives, propyl and dimethylsilyl methyl derivatives, trimethylsilyl alkyl alkoxime derivatives, deuterated derivatives; column 200-250°C depending on the <i>t_r</i> , injector 280°C, detector 300°C	416
	GC-MS	3% SE-30 on Chromosorb W, 80-100 mesh; or 3% OV-17 on Gas-Chrom Q, 100-120 mesh	Helium, 30 ml/min	MS	50 pg	Plasma	Column 230°C (SE-30), 285°C (OV-17)	
	GC, GC-MS	3% SE-30 on Chromosorb W, 80-100 mesh; 3% OV-17 on Gas-Chrom Q, 100-120 mesh	Helium, 45 ml/min	NPD, MS	1 ng/ml	Plasma	Column 225°C (SE-30), 275°C (OV-17), injector 290°C, detector 400°C; derivatization: acetylation and methylation	417

(Continued on p. 460)

TABLE 15 (continued)

Drugs separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Cytarabine + metabolites	HPLC	Aminex A-27 or A-29 (13.5 or 9.0 μ m), 65°C; or RP μ Bondapak column Co-Pell ODS Spherisorb ODS (5 μ m)	0.025 M Sodium citrate and 0.08 M sodium tetraborate buffer, 0.7 ml/min 0.01 M Potassium phosphate, 0.6 ml/min 0.05 M Phosphate buffer pH 7.0, 1.6 ml/min	UV 270 nm	2.5 ng	Plasma, urine		418
	HPLC	Partisil PKS 10/25 SCX Ultrasphere-octyl (5 μ m)	0.01 M Ammonium formate pH 4.8, 1.0 ml/min Methanol-0.01 M potassium phosphate buffer pH 7.0 (1.5:98.5), 1.6 ml/min	UV 254 nm	50 ng/ml	Plasma, CSF	1 l of 0.05 M disodium phosphate + 704 ml of 0.05 M potassium phosphate	419
	HPLC	Zorbax C ₁₈	Methanol-0.14 M sodium acetate (65:35) cont. 3 mg/l diiodine edetate, 0.9 ml/min; 28°C	Electrochemical detection	4 ng/ml (plasma); 15 ng/ml (nasal wash); 20 ng/ml (urine)	Plasma, nasal wash, urine		420
Enviroxime	HPLC	TLC	Silica gel	Multiple development: (1) <i>n</i> -butanol-acetic acid-water (4:1:2) (10–15 cm); (2), <i>n</i> -butanol-acetic acid-water (4:1:2) (3–4 cm); (3) <i>n</i> -butanol-2 M ammonium hydroxide (10:2) (to the top)	¹⁴ C Autoradiography	Urine		423
Hypoxanthine arabinoside, see Vidarabine								
Inosine pranobex (Inosiplex)	TLC	Scandium ¹ H ECD	21 ng/ml	Plasma, serum	Derivatization with chlorodifluoracetic anhydride; column 220°C, injector 230°C, detector 280°C		424	
Moroxydine	GC	2% OV-225 on Chromosorb G	Nitrogen, 30 ml/min	Scandium ¹ H ECD	21 ng/ml	Plasma, serum	Derivatization: silylation; column programmed 175°C to 240°C (8°C/min), then 270°C (serum), 230°C (urine isothermally), injector 260°C, connecting tube 240°C	425
Ribavirin (Virazole)	GC-MS	3% OV-17 or 3% SE-30 on Chromosorb W HP, 80–100 mesh	Methane, 138.3 Pa pressure	MS	10 ng/ml (serum); 0.5 μ g/ml (urine)	Serum, urine		426
	HPLC	μ Bondapak C ₁₈ (10 μ m)	5 mM Ammonium formate	UV 235 nm (220–228 nm)	20 pmol	Urine, plasma		426

Riboxamide	HPLC	(A) ODS Hypersil (5 μ m); (B) μ Bondapak C ₁₈ (10 μ m); (C) μ Bondapak CN (10 μ m); (D) Hypersil (5 μ m); anion-exchange columns: (E) Partisil 10 SAX (10 μ m); (F) μ Bondapak C ₁₈ (10 μ m) + HTAB; (G) ODS Hypersil (5 μ m) + HTAB; (H) Hypersil (5 μ m) + HTAB (5 μ m), 40°C	0.1% Sulphuric acid (pH 2.1) for A; 10 mM phosphate buffer (pH 7.0) for A-B; 10 mM phosphate buffer (pH 6.0) for F-H	UV 254 nm	40 ng/ml	Plasma	Automated column switching; 427 1 mM HTAB (hexadecyltri-methylammonium bromide)
Vidarabine, hypoxanthine arabinoside	HPLC		LiChrosorb RP-8 pH 7.2, 1.0 ml/min	Acetonitrile—5 mM pentanesulphonate buffer UV 250 nm	0.5 μ g/ml (serum or CSF); 2.5 μ g/ml (urine)	Serum, urine, CSF	428

TABLE 16
APPETITE DEPRESSANTS

Cross references between individual types of antidepressants are not supplied. Check the whole table for completeness.

Drugs separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
6-Chloro-2-(1-perazine)pyrazine	GC	3% OV-210 on Gas-Chrom Q, 100-120 mesh	Argon-methane, 50 ml/min	^{63}Ni ECD	10 ng/ml	Plasma, urine	Column 190-205°C, injector and detector by 20-30°C higher than the column Source 210°C, separator 250°C, injector 245°C	429
	GC-MS	3% OV-210 on Gas-Chrom Q, 100-120 mesh	Helium, 30 ml/min	MS				
Phendimetrazine, phenmetrazine	GC	SP-2250 on Supelcoport, 100-120 mesh	Helium, 30 ml/min		Below 10 ng/ml	Biological fluids	Oven 150°C, injector 250°C, detector 350°C; interference with diethylpropion	430
Phentermine, mephentermine	GC	12.5% Apiezon L and 2% (GEPAL) CC-380 on Chromosorb W AW (washed with 5% KOH/ methanol)	Helium, 60 ml/min	FID	ca. ng/ml	Blood, urine	Oven 140°C, injector 200°C detector 400°C	431
Amphetamine, ephedrine, mephentermine, methamphetamine, phendimetrazine, β -phenethylamine, phenmetrazine, phentermine	GC	10% Apiezon L-10% KOH on Chromosorb W (NAW), 80-100 mesh; or 3% OV-225 on Chromosorb W (AW + DMCS), 80-100 mesh		FID	1.5 $\mu\text{g}/\text{ml}$ phenmetrazine	Urine	Column 150°C, GC comparison with EMIT and RIA trifluoro- acetanilide derivatives	432, 433
Chlorphentermine, clorteramine, diethylpropion, fenturamine, mazindol, phendimetrazine, phenmetrazine, phentermine	TLC	Silica gel	(1) Ethyl acetate-methanol-water-ammonia (85:13:1:1); (2) dioxane-benzene-ammonia (35:60:5); (3) ethyl acetate-cyclohexane-ammonia (60:40:1); (4) chloroform-ammonia-acetone (95:0:1:5)		Chemical detection	Limits in $\mu\text{g}/\text{ml}$: Urine chlorphentermine 1.0, clorteramine 1.0, diethylpropion 0.1, fenturamine 0.3, mazindol 0.1, phendimetrazine 0.1, phenmetrazine 0.1, phentermine 0.1, phenmetrazine 0.5, phen- termine 1.0	Column reactions; TLC com- parison with EMIT; tables of interfering substances	434

(-)- <i>threo</i> -Chloro- citric acid	GC-MS	OV-17 on μ Pardisorb	Methane, 1.5 kg/cm ²	MS	0.1-0.6 μ g/ml	Plasma	Column 170°C, injector 300°C, interface 250°C, transfer line 250°C	436
Tiflorex + metabolite (nor.tiflorex)	GC	3% OV-17 on Gas-Chrom Q, 80-100 mesh	Nitrogen, 50 ml/min	63 Ni ECD	1 ng/ml (tiflorex); 0.5 ng/ml (metabolite)	Plasma	Derivatization with trichloro- acetyl chloride; column 210°C, injector 250°C, detector 275°C	437

TABLE 17
IMMUNOSUPPRESSIVES

Drugs separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
Azathioprine + metabolite (6-mercaptopurine)	HPLC	μ Bondapak C ₁₈ (10 μ m) for azathioprine; LiChrosorb RP-18 (10 μ m) for mercaptopurine	1.1% Acetonitrile in 0.01 M sodium acetate buffer (pH 4.0), 2 ml/min; 1% Methanol, 0.5% acetonitrile and 60 ng DTE in 0.005 M potassium phosphate buffer pH 4.0, 2 ml/min	UV 280 nm UV 325 nm	0.05 μ g/ml 5 ng/ml	Plasma		438
	HPLC	μ Bondapak C ₁₈ (10 μ m)	9% Acetonitrile in 0.01 M aq. KH_2PO_4 solution, 1.5 ml/min	UV 280 nm	2 ng/ml	Serum	6-Mercaptopurine is converted to stable derivative with N-ethyl-maleimide prior to chromatography	439
Azathioprine + metabolites (6-mercaptopurine, 6-hydroxyazathioprine, 8-hydroxy-6-mercaptopurine)	HPLC	Determination of 8-hydroxy-6-mercaptopurine: μ Bondapak C ₁₈ (10 μ m); Bondapak C ₁₈ (10 μ m)	4% Methanol and 0.5% acetic acid in 0.005 M heptanesulfonic acid pH 3.6, 1.5 ml/min	UV 325 nm		Plasma	Details see ref. 438	440
Bredinin	HPLC	LiChrosorb NH ₂ (10 μ m)	Methanol—water (30:70)	Below 0.6 μ g/ml	Blood			441
Cyclosporin A (Sandimmune)	HPLC	LiChrosorb RP-8, 72°C	0.1 M Imidazole hydrochloride buffer pH 7.0—acetonitrile (30:70), 1 ml/min	UV 280 nm	0.25 μ g/ml	Serum		442
	HPLC		(A) Water—acetonitrile—methanol (5:75:20), 1.5 ml/min; (B) water—acetonitrile—methanol (60:20:20), shape of the gradient is described	UV 210 nm	About 20 ng/ml	Plasma, urine		443
	HPLC	Supelcosil LC-8 (5 μ m) 75°C (A); segment containing cyclosporin is automatically diverted to an LC-18 Supelcosil (5 μ m), 75°C (B)	(A) 5% Acetonitrile—water (55:45), 3 ml/min; (B) 3 l acetonitrile—water (75:25), 1 ml/min	UV 202 nm	8 ng/ml (plasma); 20 ng/ml (blood)	Plasma, blood	Semiautomated HPLC	444

HPLC	Supelco LC-18 (5 μ m), 75°C	Acetonitrile—water (68:5:31:5), UV 202 nm 1.4 ml/min	130 ng/ml (blood); 110 ng/ml (plasma) ca. 100 ng/ml	Plasma, blood	445	
HPLC	Ultrasphere ODS (5 μ m), 70°C; pre- column Vydac C ₁₈ RP Ultrasphere octyl RP (5 μ m)	(A) 1% TFA in water; (B) acetonitrile; linear gradient from A→B (35:65) to (5:95), 15 min Acetonitrile—methanol—water (47:20:33), 1.5 ml/min	UV 205/215 nm 31 μ g/ml	Serum Plasma Comparison with RIA	446 447	
HPLC	Purity of cyclo- sporin. LiChrosorb RP-8 (10 μ m); or Bondapak phenyl (10 μ m), 50°C; metabolites: LiChrosorb RP-18 (10 μ m), 70°C Purification of metabolites: Amberlite XAD-2 (100–200 μ m); DEAE-Sephadex A-25, μ Bondapak- Phenyl, Porasil B (27–75 μ m)	Acetonitrile—water (6:4)	UV 210 nm 31 μ g/ml	Plasma	448	
LC		Gradient development. methanol—acetonitrile—water, 4 ml/min Water—methanol or water— methanol—0.05 M ammonium hydrogen phosphate, stepped elution with pure solvents		Urine, bile, faeces	448	
HPLC	Cyclosporin A + metabolites	Perisorb RP-8 (30–40 μ m); LiChrosorb RP-18 (5 μ m), 70°C; Ultrasphere ODS; guard column Perisorb RP-2 (30–40 μ m) μ Bondapak C ₁₈ RP, 60°C	(1) Acetonitrile—methanol— water (35:20:45); (2) aceto- nitrile—water (55:45); (3) acetonitrile—water (72:28); (4) tetrahydrofuran; (5) acetonitrile—water (90:10); (6) methanol	UV 20 ng/ml	Plasma, blood Column-switching HPLC	449
HPLC		Perisorb RP-8 (30–40 μ m); LiChrosorb RP-18 (5 μ m), 70°C; Ultrasphere ODS; guard column Perisorb RP-2 (30–40 μ m) μ Bondapak C ₁₈ RP	Methanol—water (95:5), 1 ml/min	UV 220 nm About 100 ng/ml Blood Plasma	C ₁₈ , Sample-preparation cartridges	450
HPLC				100 ng/ml	Plasma	451

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TABLE 17 (continued)

Drugs separated	Method	Stationary phase	Mobile phase or carrier gas	Detection	Sensitivity	Source	Note	Ref.
<i>erythro</i> -9-(2-Hydroxy-3-nonyl)hypoxanthine	HPLC	Ultrasphere ODS (5 μ m), 30°C	(1) Methanol-0.05 M phosphoric acid (25:75); (2) methanol-0.05 M sodium phosphate pH 7.0 (25:75); (3) methanol-0.05 M phosphoric acid (10:90); (4) Methanol-water-acetic acid (25:74:0.22); (5) methanol-water (26:74)	UV 254 nm	Urine	Also GC-MS of trimethylsilyl derivatives on 1% OV-17; column 180°C, ion source 250°C		452
	Preparative HPLC	Spherisorb ODS RT (10 μ m)	(6) Methanol-0.05 M phosphoric acid (26:74), 3 ml/min			Also preparative TLC on silica gel in seven solvent systems; liquid scintillation counting; ^{14}C derivatives		
6-Mercaptopurine	GC	3% SP 2250 DA on Supelcoport, 100-120 mesh	Helium, 20 ml/min	FID	20 ng/ml	Serum	Derivatization with trimethyl-anilinium hydroxide; column 220°C, injector and detector 300°C	453
	HPLC	Altex Ultrasphere ODS (5 μ m); pre-column Co-Pell ODS (30-38 μ m)	Acetonitrile-acetic acid-water (3.5:0.2:96.3), 1.4 ml/min	UV 322 nm	5 ng/ml	Plasma		454

3. LIST OF ABBREVIATIONS

aq.	= aqueous
conc.	= concentrated
cont.	= containing
CSF	= cerebrospinal fluid
dist.	= distilled
DTE	= dithioerythritol
ECD	= electron-capture detection
EMIT	= enzyme-multiplied immunoassay technique
FID	= flame ionization detection
GC	= gas chromatography
HPLC	= high-performance liquid chromatography
HPTLC	= high-performance thin-layer chromatography
HTAB	= hexadecyltrimethylammonium bromide
IR	= infrared
LC	= liquid chromatography
MS	= mass spectrometry
NPD	= nitrogen—phosphorus detection
PC	= paper chromatography
PIC	= PIC reagent B-7, heptanesulphonic acid containing acetic acid (PIC reagent B-7, Waters Assoc., Milford, MA, U.S.A.)
RIA	= radioimmunoassay
RP	= reversed-phase
SIM	= selected-ion monitoring
TFA	= trifluoroacetic acid
TLC	= thin-layer chromatography
<i>t</i> _R	= retention time
UV	= ultraviolet

4. SUMMARY

Some important facts about the chromatographic separation of a number of selected categories of drugs are summarized. The data refer to the chromatographic method of choice, stationary phase, mobile phase (carrier gas), detection procedure and (where available) method sensitivity. Also, fundamental instrumental parameters, namely injector, column and detector temperature, carrier gas and mobile phase flow-rate and gradient set-up are reported here. In all cases also the source material used for analysis is specified. The data are presented in table form, each table dealing with a particular category of drugs. The following categories of drugs are being dealt with: anthelmintics, antiarteriosclerotics, antibacterials, anticholinergics and cholinergics, anti-coagulants, antidiabetics, antiemetics, antimycotics, antihistamines, anti-malarials, antiparasitics, antiparkinsonics, antitussives, antiulcer drugs, anti-viral compounds, appetite depressants and immunosuppressives.

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